

# Screening and identification of lactic acid bacteria strains with high acid-producing from traditional fermented yak yogurt

Xiaoyong Chen<sup>1,2,3,#</sup>, Xin Zhao<sup>3,4,#</sup>, Yu Qian<sup>3,4</sup>, Jian Li<sup>5,6</sup>, Lianhong Chen<sup>5,6</sup>, Juan Chen<sup>6</sup>, Yu Zhang<sup>1,2</sup> and Huayi Suo<sup>1,2,a</sup>

<sup>1</sup>College of Food Science, Southwest University, 400715, Chongqing, China

<sup>2</sup>Chongqing Engineering Research Center of Regional Food, 400715, Chongqing, China

<sup>3</sup>Chongqing Collaborative Innovation Center for Functional Food, Chongqing University of Education, 400067, Chongqing, China

<sup>4</sup>Department of Biological and Chemical Engineering, Chongqing University of Education, 400067, Chongqing, China

<sup>5</sup>Institute of Qinghai-Tibetan Plateau, Southwest University for Nationalities, 610041, Chengdu, China

<sup>6</sup>College of Life Science and Technology, Southwest University for Nationalities, 610041, Chengdu, China

**Abstract.** A total of 57 strains of lactic acid bacteria (LAB) were isolated and purified from traditional fermented Yak Yogurt in Hongyuan-Sichuan and Yangbajing-Tibet. The strains with high acid-produced were screened by soluble calcium circle and titratable acidity determination. The five strains, 7-1, 22-1, 28-1, 34-1 and 62-1, possessed the high acid-producing and the value of titratable acidity is 196.2, 191.1, 192.2, 194.8 and 200.2 T respectively. Based on 16S rDNA sequence analysis, 22-1 was identified as *Lactococcus lactis* subsp. *lactis*, 28-1 as *Lactobacillus casei*, 34-1 as *Lactobacillus fermentum*, 7-1 and 62-1 as *Enterococcus durans*. This study could provide the evidence for researching fermentation strains to improve yogurt quality.

## 1 Introduction

Lactic acid bacteria refer to a group of bacteria with the capacity to ferment carbohydrate to generate lactic acid. It is a main fermentative strain to ferment traditional fermented foods like dairy product, pickle and fermented sausage etc. Lactic acid bacteria generates flavor substances such as micromolecule aldehyde, acid and ester etc. through fermenting saccharides to promote the formation of traditional fermented food's flavor, which plays a great importance in fermented food [1, 2]. In the fermentation process of dairy product, the acid-producing ability of lactic acid bacteria is a key factor to measure the quality of dairy product [3]. Bian Lihong and others [4] researched and found that the acid-producing ability of lactic acid bacteria in different brands of plain yogurt is closely related to the

---

<sup>a</sup> Corresponding author: birget@swu.edu.cn, <sup>#</sup> Co-first author

This work was partly supported by Special Fund for Agro-scientific Research in the Public Interest (201203009), Special Fund of Chongqing People's Livelihood and Technology Innovation (cstc2015shmszx80021) and Fundamental Research Funds for the Central Universities(XDJK2016E109).

separation of curd and whey in yogurt. In addition, the lactic acid bacteria with a high acid-producing ability can not only shorten curd time and enhance productivity but also perfect yoghurt's texture and improve the quality of yoghurt etc. Consequently, to select the lactic acid bacteria with a high acid-producing ability has an important significance for improving the quality of yoghurt. At present, to select the lactic acid bacteria with a high acid-producing ability among traditional fermented foods has gotten a wide attention from many domestic and overseas scholars. Zhang Heping etc. [5] researched the characteristics of acid production for the lactic acid bacteria separated from natural fermented dairy product, from which a *Streptococcus thermophilus* with short curd time, high rate of acid production and weak post-acidification ability is obtained. Ma Yuxiao etc. [6] get three lactic acid bacteria with high curd speed, high acid-producing ability and fine curd quality through the screening from dzo yoghurt. In addition, Carafa etc. [7] researched the experiment on glycometabolism, acidification ability and acid and bile salt tolerance etc. of lactic acid bacteria in natural fermented cheese of Alps, the selected lactic acid bacteria from which can be applied in the production of fermented dairy product. The good curd quality in traditional yak yoghurt has shown that there are lactic acid bacteria with a high performance. The special growing environment and the narrow distribution range of yak have limited the research of lactic acid bacteria in traditional fermented yak yogurt.

Our country is the headstream of yak in the world. Yak resource is very rich in Qinghai-Tibet Plateau, where herdsman adopts traditional handicraft and natural fermentation method to make yak yogurt [8, 9]. It is an indispensable necessity in herdsman's daily life, as well as a traditional featured delicious food in this area. The unique geographical environment, climatic condition, Tibetan herdsman's scattered community and nomadic pasture pattern have together formed the special resource pool of lactic acid bacteria in Qinghai-Tibet Plateau, which offers a guarantee for selecting the lactic acid bacteria with high fermentative capability. This research screens the lactic acid bacteria in traditional fermented yak yogurt and evaluates its acid-producing ability to provide the reference for better using fine strains and improving the quality research of traditional fermented yak yogurt.

## **2 Material and method**

### **2.1 Reagents**

#### *2.1.1 Bacterial source*

Fifty seven lactic acid bacteria strains separated from the traditional yak yogurt in Hongyuan of Sichuan and Yangpachen of Tibet are kept in store by College of Food Science, Southwest University.

#### *2.1.2 Reagent*

MRS broth, agar from Beijing Solarbio Science and Technology Ltd. Sodium hydroxide, calcium carbonate analytically pure from Chengdu Kelong Chemical Reagent Factor.  $\lambda$ DNA/HindIII, 6 $\times$ DNA Loading Buffer, 100 bp DNA Ladder, 2 $\times$ Taq PCR MasterMix, bacterial genome DNA extraction kit (DP302) from Tianjin Biotech (Beijing) Co., Ltd. forward primer 27F (5-AGAGTTTGATCCTGGCTCA-3), reverse primer 1495R (5-CTACGGCTACCTTGTTACGA-3) compounded by Sangon Biotech (Shanghai) Co., Ltd.

### **2.2 Instrument and equipment**

Storm Gradient PCR, S1000 Thermal Cycler and Mini-Sub Cell GT from American Bio-Rad Company, Centrifugal machine 8510 from Germany Eppendorf Company, Water-Jacket Thermostatic Constant Incubator, GHP-9160 from Shanghai Keelrein Scientific Instrument Co., Ltd. Clean bench,

SW-CJ-2F from Suzhou Antai Airtech Co., Ltd. pH meter, PHS-3C from INESA Analytical Instrument Co., Ltd.

## **2.3 Methodology**

### *2.3.1 Preliminary screening of lactic acid bacteria with high acid-producing ability*

In clean bench, add 1.5% sterile CaCO<sub>3</sub> into the sterile MRS medium. Then shake to make CaCO<sub>3</sub> well mixed. In this process, bubble should be avoided. When it is cooled to about 46°C, pour it into the plate immediately. After solidification, it is as standby application. Select an appropriate dilution of bacterial liquid to make spread plate cultivation at 37°C for 24h. Finally, make the preliminary judgment for the acid-producing ability of experimental bacteria through the size of CaCO<sub>3</sub> lysis zone and bacterial colony to select the bacterial strain with an obvious transparent zone as the alternative bacterium with high acid-producing ability.

### *2.3.2 Determination of acidity for lactic acid bacteria with high acid-producing ability*

Inoculate the cultivated bacterial liquid into MRS fluid medium as per 2% inoculum size. Measure the titration acidity after cultivating for 48h at 37°C. Take 10 mL nutrient solution into 150 mL erlenmeyer flask. Then add 20 mL water after boiled and cooled and 2 drops of phenolphthalein indicators to make well mixed. Use acidometer to indicate end point and 0.1 mol/L NaOH standard solution to titrate to pH, 8.2. Record the volume of NaOH standard solution consumed. Finally calculate the acidity of nutrient solution, which will be indicated by Thorner degrees (°T) [10].

### *2.3.3 Analysis on identification and phylogenetic tree of lactic acid bacteria with high acid-producing ability*

Preparation of DNA template: extract genome DNA (template) of lactic acid bacteria with high acid-producing ability by the method in the specification of DNA extraction kit. Use 0.7% agarose gel electrophoresis to make the test.

PCR amplification: PCR reaction system is 25  $\mu$ L. Template is 1  $\mu$ L. Primers, 27F and 1495R are 1  $\mu$ L, 2 $\times$ Taq PCR MasterMix 12.5  $\mu$ L respectively, complemented to 25  $\mu$ L through adding sterile ultrapure water. PCR reaction conditions include 94°C 5min, 94°C 40s, 55°C 40s and 72°C 1min, totaling 35 cycles as well as 72°C 10 min. After the reaction is finished, take 5  $\mu$ L PCR products to make the test through 1.5% agarose gel electrophoresis.

Sequencing comparison: send PCR products to BGI Science and Technology Ltd. to make sequencing. The sequencing data is submitted to GenBank of NCBI. Use Blast program to search the sequence with maximal homology, and finally determine the species of lactic acid bacteria with high acid-producing ability.

Analysis on phylogenetic tree: take 10 kinds of 10 lactic acid bacteria's gene sequences in the same segments from GeneBank database. Use Alignment program in Clustalx1.83 software to make multi-sequencing matching comparison for the tested sequences and the reference sequences obtained from BLAST searching. Then use Kimura 2-parameter mode in MEGA5.0 software to calculate genetic distance. The phylogenetic tree of homologous sequence to separate lactic acid bacteria is constructed by neighbor-joining method. Each branch of phylogenetic tree adopts bootstrap method to make confidence coefficient test. The bootstrap data set is 1000[11-13].

## **3 Results and analysis**

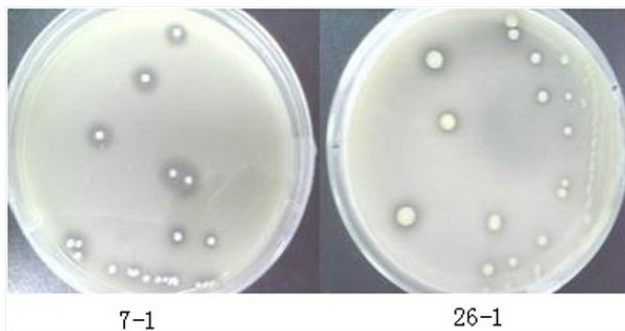
### **3.1 Preliminary screening results of high acid produced lactic acid bacteria**

Table 1 and Figure 1 show that the calcium-dissolving zones of 17 lactic acid bacteria are found not obvious. 30 lactic acid bacteria can generate calcium-dissolving zone. But the transparent zones are different to some extent for different bacterial strains.

**Table 1.** The situation of lactic acid bacteria produced transparent circle.

Serial No.	No.	Transparent circle size	Serial No.	No.	Transparent circle size	Serial No.	No.	Transparent circle size
1	27	+	20	4-1	+	39	25-1-1	++
2	31	-	21	4-2-2	++	40	25-1-2	++
3	32-2	++	22	5-1	++	41	25-2	+
4	37	-	23	6-1	+	42	26-1	-
5	38-2	-	24	6-2	+	43	27	++
6	41-1	-	25	7-1	++	44	27-2	++
7	43-2	-	26	7-2	++	45	28-1	++
8	54	++	27	12-1	++	46	32-1-1	++
9	55	-	28	16	+	47	32-1-2	++
10	57-2	+	29	17-2	++	48	34-1	++
11	58	-	30	18-1-1	++	49	40	++
12	60	-	31	18-1-2	++	50	41-1	++
13	64	+	32	18-2	+	51	41-2	+
14	71	-	33	18-3	++	52	42-1	+
15	77	++	34	19-1	++	53	53-1	++
16	88	++	35	21-1	+	54	53-2	+
17	89	-	36	21-2	++	55	53-3	+
18	3-1-1	++	37	22-1	++	56	62-1	++
19	3-1-2	+	38	23-1	++	57	67-1	+

Note: "++", strong transparent circle, "+", weak transparent circle, "-", no transparent circle.



**Figure 1.** The lactic acid bacteria produced transparent circle in MRS calcium carbonate medium.

### 3.2 Acidity determination results of high acid produced lactic acid bacteria

Table 2 shows that the acid-producing range of experimental bacterial strain is from 69.2 to 200.2<sup>o</sup>T, among which the acid-producing contents of No.7-1, 22-1, 28-1, 34-1 and 62-1 bacterial strains are higher. The titration acidities are 196.2, 191.1, 192.2, 194.8 and 200.2 o T respectively. The acid-producing contents of experimental bacterial strains researched by Liu Zhenming etc. [14] are from 64.4<sup>o</sup>T to 294.2<sup>o</sup>T, and the ones researched by Zhang Lanwei etc. [15] from 44.35<sup>o</sup>T to 57.65<sup>o</sup>T. Compared with the screening results made by other scholars, the lactic acid bacteria with bacterial strains No. 7-1, 22-1, 28-1, 34-1 and 62-1 have a higher acid-producing ability. Under the same

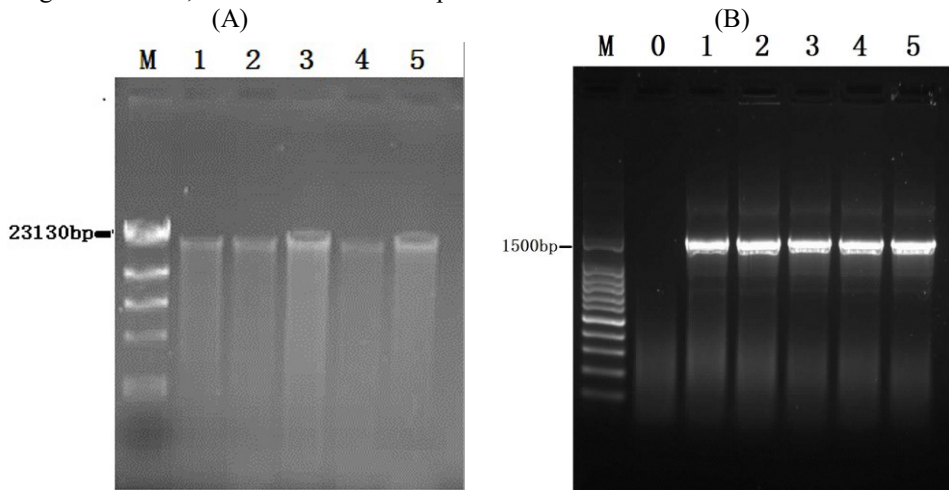
culture medium and condition, the different acid-producing ability of different strains shows that difference exists between the acid-producing ability of bacterial strain metabolism's carbohydrate and its acid resistance.

**Table 2.** Acidity determination results of high acid produced lactic acid bacteria after screening.

Serial No.	No.	Acidity (°T)	Serial No.	No.	Acidity (°T)	Serial No.	No.	Transparent circle size
1	32-2	91.9±0.36	11	17-2	128.8±1.08	21	27	103.3±0.53
2	54	91.8±0.53	12	18-1-1	104.8±0.26	22	27-2	69.2±0.79
3	77	98.0±0.20	13	18-1-2	105.9±0.35	23	28-1	192.2±0.30
4	88	87.8±0.26	14	18-3	104.0±0.26	24	32-1-1	129.7±0.36
5	3-1-1	86.8±0.46	15	19-1	103.2±0.30	25	32-1-2	104.0±0.26
6	4-2-2	102.9±0.36	16	21-2	110.2±0.36	26	34-1	194.8±0.79
7	5-1	134.2±0.89	17	22-1	191.1±0.20	27	40	100.4±0.44
8	7-1	196.2±0.26	18	23-1	177.8±0.23	28	41-1	128.0±0.26
9	7-2	145.3±0.36	19	25-1-1	103.0±0.70	29	53-1	94.2±0.44
10	12-1	122.4±0.30	20	25-1-2	153.8±0.26	30	62-1	200.2±0.46

### 3.3 Identification of high acid produced lactic acid bacteria

Figure 2A shows that genome DNA of bacterial strain is in around 23130bp, strip neat and luminosity obvious, which means the extraction effect of genome DNA is good. The target fragment after PCR amplification gets close to 1500bp (Figure 2B), no tailing phenomena in strip, clear and bright, and no strip in negative control, which means PCR amplification succeeds with a better effect.



**Figure 2.** Agarose gel electrophoresis of genomic DNA (A) and PCR products (B).

Traditional fermented yak yogurt is a dairy product with unique flavor, co-fermented by complex microflora taking actic acid bacteria and saccharomycetes as main bacteria and microorganism. At present, it is found through research that its fermentation microorganism [16] is mainly composed by *Enterococcus faecium* and *Enterococcus durans* in *Enterococcus*, *Lactobacillus casei*, *Lactobacillus plantarum*, *Lactobacillus bulgaricus*, *Lactobacillus fermentium* and *Lactobacillus helveticus* in *Lactobacillus* and *Leuconostoc mesenteroides* subsp. *mesenteroides* as well as *Streptococcus thermophilus* etc. Traditional fermented yak yogurt is characterized by high acidity. The acid generally [17, 18] can reach to 198.7°T. The research finds various lactic acid bacteria including *Enterococcus faecium*, *Enterococcus durans*, *Lactobacillus casei*, *Lactobacillus paracasei*, *Lactobacillus fermentium* and *Lactococcus lactis* as well as *Lactococcus lactis* subsp. *lactis* and *Streptococcus thermophilus* etc. contribute to the formation of yak yogurt's acid and flavor. Table 3 shows the experiment also



## 4 Conclusion

In this research, the lactic acid bacteria with high acid-producing ability in traditional fermented yak yogurt is screened and selected by calcium-dissolving zone method and through determination of titration acidity. The research shows the abilities of different bacterial strains in yak yogurt to generate calcium-dissolving zone are different. The acid-producing abilities are also different. The acidity is between 69.2 and 200.2 o T. Among them, the bacterial strains with No. 7-1, 22-1, 28-1, 34-1 and the bacterial strain with No. 62-1 have a higher acid-producing ability. After cultivating for 48h at 37<sup>o</sup>C, titration acidities are 196.2, 191.1, 192.2, 194.8 and 200.2 o T respectively. No. 22-1 bacterial strain is identified as *Lactococcus lactis* subsp. *lactis*, No. 28-1 bacterial strain as *Lactobacillus fermentum*, and the bacterial strains with No. 7-1 and 62-1 as *Enterococcus durans* through 16S rDNA sequence analysis. Although *Enterococcus durans* have not been recorded in the list of bacteria used in our country's food industry, it is an important composition in microflora of traditional fermented yak yogurt. The experimental result shows that it has a high acid-producing ability, and playing an important role in the formation of flavor and quality of traditional fermented yak yogurt.

## References

1. T.Q. Zhang, Z.N. Yang, B.H. Kong, Food Sci 637-642, **29** (2008).
2. X. Xin, G. Song, X.H. Zhou, J.W. Zhao, L. Chen, Q.P. Ge, W.X. Ping, J. Chinese Inst. Food Sci. Technol. 202-207, **14** (2014).
3. H. Qin, X. Liang, W.B. Zhang, Y. Zhang, L. Mi, Food Sci. 241-246, **34** (2013).
4. L.H. Bian, Y. Wang, Y.T. Zhang, L.N. Qu, Guangdong Agri. Sci. 128-130, **39** (2012).
5. H.X. Chen, L.L. De, Y. Ren, D.L. Zhang, Y.R. Yang, W.J. Liu, H.P. Zhang, J. Dairy Sci. Technol. 1-5, **2015** (2015).
6. H. Tian, X.P. Zhang, Y.L. Yan, Food Sci. 152-156, **31** (2010).
7. I. Carafa, T. Nardin, R. Larcher, R. Viola, K. Tuohy, E. Franciosi, Food Microbiol. 123-132, **48** (2015).
8. C.S. Wu, C. Shu, J. Li, Y. Qian, H.Y. Suo, Food Ind. 129-133, **33** (2012).
9. Y.W. Wang, C.M. Zhang, H.Y. Suo, H. Yue, J. Li, C. Tang, Sci. Technol. Food Ind. 184-187, **35** (2014).
10. C.X. Liu, Jiangsu J. Prev. Med. 69-70, **19** (208).
11. N.A. El-Naggar, S.A. Haroun, E.A. Oweis, A.A. Sherief, Prep. Biochem. Biotechnol. 712-729, **45** (2015).
12. K. Tamura, D. Peterson, N. Peterson, G. Stecher, M. Nei, S. Kumar, Mol Biol Evol. 2731-2739, **28** (2011).
13. S. Ennahar, Y. Cai, Y. Fujita, Appl. Environ. Microbiol. 444-451, **69** (2003).
14. Z.M. Liu, Y.Y. Wang, J. Dairy Sci. Technol. 169-172, **33** (2010).
15. C.L. Ma, L.W. Zhang, Sci. Technol. Food Ind. 189-190, **31** (2010).
16. L.V. McFarland, Clin. Infect. Dis. 85-90, **60** (2015).
17. Z. Luo, Y.G. Shi, L. Yu, B.Z. Han, China Brew. 40-41, **2005** (2005).
18. J. Xu, Y.Y. Yun, W.Y. Zhang, Y.D. Shao, B. Menghe, H.P. Zhang, China Dairy Ind. 23-27, **34** (2006).