

Comparison of the effect of 8 closures in controlled industrial conditions on the shelf life of a red wine

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Abstract. Aims: The management of O₂, CO₂ and SO₂ at bottling and the choice of the closure are two key factors of the shelf life of wine in bottles before bringing them to market. The impact of four screw caps, two synthetic and two technical corks was evaluated on a red wine of Merlot/Tannat.

Methods and results: Analytical monitoring (O₂, CO₂, SO₂, aphyrometric pressure, L*, a*, b*) was carried out during 538 days of storage at 20 °C. Two sensory analyses at 10 and 17 months completed the study.

The wine was bottled with an average total oxygen content of 2 mg/L. The heterogeneity intra and inter procedure was controlled, including for the dissolved carbon dioxide content.

Conclusion: Unlike closures with highest OTR, the two technical corks and the two screw caps with Saranex seal, harboring the lowest OTR, matched with the wines exhibiting a low total O₂ content at equilibrium (from 4th to 18th month), with more free SO₂ and less changed colour. However this OTR gradient (5 to 67 µg/d) observed through the physicochemical analyses was not necessarily confirmed by both sensory analyses performed.

Significance and impact of study: This study puts into perspective the impact of OTR closure on sensory characteristics evolution of wine consumed during the first two years, especially when the total oxygen at bottling exceeds 1.5 mg/L.

1. Introduction

Oxygen is one of the main factors for wine's evolution. At bottling, oxygen captured in the headspace (HSO) and dissolved in the wine (DO) must be reduced as much as possible. The Oxygen Transmission Rate (OTR) of stopper regulates the transfer of oxygen inside the bottle after bottling. The management of O₂, CO₂ and SO₂ at filling and the choice of stopper are the key factors of the shelf life, manageable by the conditioner before bringing bottles to market.

The oxygen ingresses during and post bottling lead to the decrease of sulfites. In wine, the reaction between O₂ and SO₂ is extremely slow [1]. Sulfites react with the products of wine oxidation and in particular with hydrogen peroxide, product of the oxidation of phenolic compounds [2,3]. The wine becomes more sensitive to oxidation and ages faster. Godden et al. [4] highlighted a critical concentration of free SO₂ of 10 mg/L below which a Semillon wine is perceived as substantially affected by oxidised aroma. For red wines, controlled oxygen ingresses are necessary and variable according to the expected quality before and after bottling especially to avoid the reduction [5,6].

The commercial choice between stopper (natural, technical or synthetic) and screw cap has a direct impact on the volume and the inerting process of the headspace as well as the OTR of closure. The volume and the management technology of headspace (vacuum, gas sparging, snowdrop) explain that the quantity of oxygen

trapped in the headspace can vary from 0.4 to 3.6 mg per bottle as measured by Vidal and Moutounet [7]. The bottling line audits outlined by O'Brien et al. [8] confirm this broad range of oxygen amount. Kontoudakis et al. [9] showed that stopper type significantly affected the HSO content. The volume of the headspace of a corked bottle is significantly lower than this of a capped bottle, but on the other hand the cork releases a portion of the oxygen trapped in its own structure due to the compression of the stopper in the bottleneck [10].

Regarding the bottle position, there is no consensus to date on an effect on oxygen mass transfer through the closure and wine aging over time, even if in theory, the oxygen diffusion coefficient through the closure into wine is smaller than into the headspace. Mas et al. [11] concluded that white and red wines were best preserved when bottles were stored horizontally rather than vertically. Puech et al. [12] on rosé and red wines and Skouroumounis et al. [13] on white wines found no significant differences. Godden et al. [4] concluded that for several closures upright storage tended to accelerate loss of SO₂ from a Semillon wine, but in many cases this effect was marginal.

The principal methods for determination of OTR used for wine closures are the coulometric method by Mocon Oxtran and nitrogen flushing of the inner face of the cell [14], the method by permeability meter and pressure difference between the two faces of stopper [15], the luminescence method on corked or capped bottles filled with nitrogen [16] or deoxygenated acid water [17] and

Table 1. Description of the 8 closures. ¹ PVDC: Polyvinylidene chloride. ² OTR: Oxygen Transmission Rate in mg/day/closure given by manufacturer. ³ OTR, included release, calculated on 1 year. ⁴ OTR, included estimated release of 1.5 mg calculated on 1 year.

Code	Type	Length mm	Diameter mm	Weight g	Density kg/m ³	OTR ² mg/d	Method of OTR measurement
B1	Synthetic stopper	42	22.5	4.8	285	0.014 ³	Luminescence
B2	Technical stopper	44	24.5	5.6	270	0.005 ⁴	Coulometry
B3	Technical stopper	44	24.0	5.7	280	0.005 ³	Colorimetry
B4	Synthetic stopper	42	22.0	7.6	488	0.047	Coulometry
C1	Screw caps + Saranex seal		31.5			0.005	
C2	Screw caps + Saranex seal		31.5			0.005	
C3	Screw cap + seal without PVDC ¹		31.5			0.067	
C4	Screw cap + seal without PVDC		31.5			0.007	

the colorimetric method with indigo carmine [18]. The Mocon Oxtran technology is by far the most commonly used in the packaging industry. But when applied to the bottle/closure system, it cannot mimic exactly the configuration of storage where the closure is wet with wine or in contact with water vapor saturated headspace in case of vertical storage. Moreover, another major drawback of this method is the long time required to reach the steady state of oxygen ingress through the closure when 40 mm long stoppers are tested [19]. These reasons explain why manufacturers use also methods with operating conditions closer to enological reality and which better integrate the desorption of oxygen by the stopper mainly during the first month as luminescence and colorimetric methods [16,20].

However, whatever the used method, the OTR range of natural corks is roughly intermediate between this of screw cap and technical stoppers and this of synthetic stoppers, but with a greater heterogeneity by comparison with industrial stoppers [15, 18, 21, 22].

Logically, many studies have shown the impact of OTR on the quality of wines. Also the higher oxygen permeability it is, the higher are decrease in the SO₂ level, increase of the absorbance at 420 nm and premature emergence of oxidised aroma for white wines [4, 13, 23–25]. Conversely, screw caps are cited by the majority of the previous articles as closures for which sensorial reduction notes are most frequent due to their low OTR.

Generally speaking, the red wine behaved in a similar way to the white wine, but thanks to its higher phenolic compounds content, it was much more resistant to oxidation [11] but also sensible to reduction when oxygen ingresses post-bottling are insufficient [5,6], in particular with screw caps [26].

Thus, for entry and mid-range wines, synthetic or technical stoppers and aluminum caps usually supersede natural corks.

Based on these findings, an experimental protocol was set up in order to answer the following questions. What is the impact of these closures on the shelf life of a red wine bottled at an industrially achievable Total Oxygen content (TO₂) and intended to be drunk within two years? Are there differences between stoppers and screw caps? Which physicochemical and sensory characteristics are influenced significantly by the oxygen permeability of the closure? To the best of knowledge, this is one of first studies based on the relationship between OTR and the consumptions of oxygen and sulfites under industrial conditions controlled by dissolved gases and sulfites.

2. Material and methods

2.1. Experiment

A 2013 IGP Côtes de Gascogne red wine (70% Merlot + 30% Tannat) was bottled on 26 June 2015 at INRA Pech-Rouge bottling facility (Gruissan, France) in 75 cL flint glass Bordeaux bottles at targeted levels of O₂, CO₂ and SO₂ and with a weak dispersion. Two synthetic stoppers (B1, B4), two technical stoppers (B2, B3), two screw caps with Saranex seal (C1, C2) and two screw caps with seal without Polyvinylidene chloride (PVDC) were tested (Table 1). The two types of closure and the different length of stoppers (42 or 44 mm) led to different headspace volumes and inerting processes. The target TO₂ in bottle was set at 1.5 mg/bt (2 mg/L), a value reasonably achievable at the industrially level whatever the closure used.

After bottling, bottles were stored upright in the dark in a thermostatically controlled room at 19.9 ± 0.5 °C, with 67.2 ± 15.8 %HR (monitored, but not controlled). Both destructive and non-destructive physicochemical analyses were carried out on several dates spread over 538 days after bottling. An expert jury performed sensory analysis at 10th and 17th months.

Given the small thickness of the seal, the oxygen release of screw caps is negligible as shown by Vidal et al. [17]. For stoppers, as discussed in the introduction, manufacturers generally prefer the luminescence and colorimetric methods of OTR measurement to better quantify the higher release of oxygen by stopper at the beginning of storage which significantly increases the OTR. The coulometry tends to undervalue this phenomenon and gives an OTR of 0.009 mg/d lower than the luminescence method for B1 (0.014 mg/d). The OTR of B2 by coulometry is enhanced by an estimated release of 1.5 mg. For the stoppers B3 and B4, manufacturers have provided OTR value obtained by a single method presented in Table 1.

2.2. Bottles

Cork bottles: OI (Villeurbanne, France), standard 75cL BD CAR II LG, unfilled level 63 mm; screw cap bottles: OI, standard 75cL BD CAR II LG BVS, unfilled level 45 mm.

2.3. Analyse of wine just after bottling

12.9% vol.; sugar 2.6 g/L; TA 3.33 g H₂SO₄/L; VA 0.43 g H₂SO₄/L; pH 3.49; free SO₂ 27 ± 0 mg/L; total SO₂

68 ± 0 mg/L; CO₂ 325 ± 15 mg/L; L* 67.21; a* 33.89; b* 8.91; A₄₂₀ 2.532; A₅₂₀ 3.379; A₆₂₀ 0.633, Total Phenol Index=49.

2.4. Bottling

The INRA Pech-Rouge bottling line for experimental wines allowed the control and management of dissolved gases on the three elements of the chain:

- a filtration skid (RS IW, Tübingen, Germany) with preparation tank (105 L), prefiltration (1 μm) and final filtration (0.65 μm),
- a single head filler MTB 1/1 (Perrier, Le Cheylar, France) with or without neutral gas flushing of filler tank (46 L) and bottles before filling,
- a single head corking machine Gemini R (Arol, Canelli, Italy) with coupling vacuum and inert gas (N₂ for this study) in several cycles before corking in order to reduce the oxygen amount of the headspace.

This line achieves homogeneous bottling of small volumes of wines with very low variations in TO₂ and dissolved CO₂ [27,28].

Four batches of 100 L were required for the filling of the 450 bottles of the study. The preparation tank was filled by gravity with the starting tank of red wine. The wine was sparged with N₂ gas using a porous injector bolted to the bottom of the preparation tank until DO reached 0.15 mg/L then adjusted to 300 mg/L of dissolved carbon dioxide (DCO₂) by sparging with CO₂ gas.

The circuit was purged with N₂ from the outlet of preparation tank to head filler machine. Wine was forced into the circuit by N₂ to the filler tank through the filtration skid using overpressure of 100 kPa applied to the top of preparation tank. Bottles were blanketed before filling. A slight depression of 8 kPa assisted the filling height adjustment.

Filled cork bottles were sealed by the single head corking machine. 2 combined cycles of N₂ (1 s) followed by vacuum 75 kPa (1 s) were performed. Filled screw cap bottles were crimped by a single head capping machine Galaxy (Costral, Riquewih, France) without inerting of headspace and cap. The unscrewing torque has been checked for the four kinds of screw cap bottles with Orbis 6 Nm digital torque tester (Mecmesin, Slinfold, England).

Since the DO has been set to a low level for all procedures, the TO₂ target value of 1.5 mg/bt was reached thanks to the management of the headspace according to the type of bottle and its unfilled level.

2.5. Physicochemical analyses

On line O₂ monitoring was performed using a PreSens luminescent probe and PSt3 O₂-sensitive optical spots (PreSens Precision Sensing GmbH, Regensburg, Germany) integrated at four checkpoints on the bottling line and at the top and bottom of the preparation and filler tanks. DCO₂ was monitored by sampling in the preparation tank using a Carbodoseur (Dujardin-Salleron laboratories, Noizay, France).

The following destructive analyses were performed at T0, 1, 4, 8, 12 and 18 months: aphrometric pressure (simplified aphrometer for still wines, Ligapal,

Table 2. Sensory attributes selected and composition of their reference standards. ¹ Attributes added at 17 months.

Sensory cluster	Attribute	Reference standard
Visual	Colour intensity	
Olfactory	Amylic	Isoamyl acetate
	Animal (leather)	4-Ethylphenol
	Cooked red fruits	Red fruits jam
Gustatory	Dry wood (dust) ¹	Unheated wood powder
	Pastry (vanilla, caramel)	Caramel syrup
Gustatory	Pepper	Black pepper
	Astringency	Grape stem tannin extract
	Bitterness	Caffeine
	Sourness	Tartaric acid
	Sweetness ¹	Grape sugar
	Alcohol ¹	Absolute ethanol

Cormontreil, France); free and total SO₂ (potentiometric titration, Titromatic, Crison Instruments, Alella, Spain); ΔEab* (spectrophotometer CM3600d, Konica Minolta, Roissy CDG, France, standard illuminant D65, 10° standard observer). Non-destructive monitoring was conducted every month from T0 to 18 months for unfilled level (set square for wine bottle), calculation of the volume of headspace, dissolved and gaseous O₂ (luminescence with PreSens PSt3 glued spots inside bottles) and dissolved CO₂ (laser spectroscopy, Lsensor CO₂, FT System, Padova, Italy). For destructive and non-destructive chemical analyses, three repetitions were performed by parameter / procedure / date.

Twelve bottles capped with C4 screw cap were stored at 7 °C (C4 7 °C). These bottles were used for SO₂ analyses at 243, 370 and 532 days.

2.6. Sensory analyses

Descriptive quantitative analysis was conducted by an expert sensory panel (22 judges), selected on the basis of their sensory performances and interest (ISO 8586-2, 1994), and trained to descriptive sensory analysis of wines. At the first step of generating vocabulary, the jury selected attributes by consensus, to describe the samples. The Table 2 presents the finally selected attributes. Then the panelists were trained to understand and consistently use attributes and familiarized with the product space. Finally, the judges rated each attribute on an unstructured linear scale from “low” to “high”. For olfactory and taste analyses, wines were evaluated in duplicate, in monadic service, according to a random order (Latin square) minimizing carry-over effects, in black tulip-shaped glasses to ensure that visual perceptions did not influence olfactory and taste analyses, between 17 and 19 °C. Following for visual attributes, samples were evaluated in comparative service, in 215 ml wine glasses, with “normal daylight” illumination.

Sensory data were converted into marks from 0 to 10 by Fizz Software version 2.40 A (Biosystemes, Couternon, France).

2.7. Statistical analyses

Statistical analyses were performed using XLSTAT software version 2014 (Addinsoft, Paris, France).

Table 3. Contents of O₂ at bottling and TO₂ at equilibrium. HSO: Headspace Oxygen; DO: Dissolved Oxygen; TO₂: Total Oxygen; mg/bt: mg/bottle. ¹ TO₂ at equilibrium: average TO₂ between 90th and 532th day. Averages and standard deviations are based on 3 bottles per procedure.

Type	Code	T0			TO ₂ equilibrium mg/bt ¹
		HSO mg/bt	DO mg/bt	TO ₂ mg/bt	
Stopper	B1	1.08 ± 0.00	0.15 ± 0.06	1.22 ± 0.06	0.06 ± 0.02
	B2	1.19 ± 0.12	0.19 ± 0.05	1.38 ± 0.09	0.04 ± 0.01
	B3	1.03 ± 0.07	0.30 ± 0.19	1.34 ± 0.17	0.05 ± 0.01
	B4	0.99 ± 0.07	0.25 ± 0.05	1.24 ± 0.12	0.11 ± 0.06
Screw cap	C1	1.43 ± 0.21	0.19 ± 0.03	1.62 ± 0.18	0.04 ± 0.01
	C2	1.46 ± 0.17	0.18 ± 0.03	1.64 ± 0.17	0.05 ± 0.01
	C3	1.61 ± 0.25	0.15 ± 0.03	1.75 ± 0.28	0.23 ± 0.03
	C4	1.54 ± 0.07	0.20 ± 0.09	1.74 ± 0.13	0.07 ± 0.02
Stoppers averages		1.07 ± 0.06 (10.4 ± 0.6 %v/v)	0.25 ± 0.09	1.30 ± 0.11	
Screw caps averages		1.51 ± 0.17 (7.9 ± 0.9 %v/v)	0.18 ± 0.05	1.69 ± 0.19	

For sensory data, after confirming jury good performances (repeatability, consensus and discrimination), results were processed by analysis of variance (2 factors: judge and wine). When significant differences were revealed ($p < 0.05$), mean intensities were compared using the Tukey (HSD) multiple comparison test.

The results of free - total SO₂ and consumption ratios of SO₂/O₂ were processed by analysis of variance (1 factor: closure). When significant differences were revealed ($p < 0.05$), analytical parameters were compared using the Tukey (HSD) multiple comparison test.

The treatment allowed to classify the different wines in several distinguishes groups (A, B, C, D, E). The average values and the several groups are given in this paper.

3. Results and discussion

3.1. Industrial bottling parameters

The unscrewing torque of capped bottles tested after crimping was on average 16 ± 1 lbf/inch for all four screw caps procedures. The unfilled level which determines the volume of the headspace was on average 63 ± 2 mm for the stoppers and 45 ± 2 mm for screw caps (net of seal thickness) from T0 to 538 d. Aphrometric pressure fluctuated between -160 and 200 kPa from T0 to 538 d. The monitoring of these three parameters was in accordance with usual technical recommendations.

3.2. Dissolved gases

Just after bottling, 86% of TO₂ (TO₂ = HSO + DO) was located in the headspace of bottle. The average TO₂ was 1.5 mg/bt (2 mg/L). Heterogeneity remained limited because the highest intraprocedural standard deviation was 0.28 mg/bt (Table 3) and the maximum intraprocedural deviation was 0.53 mg/bt (between B1 and C3). As already described several times [29–31], both oxygen in the headspace and dissolved in the wine were consumed. This decrease was not linear because 90% of initial TO₂ was consumed after 35 days. From 48th day, TO₂ was less than 0.10 mg/bt, except for B4 and C3. These procedures had the most variable and highest TO₂ content, mainly

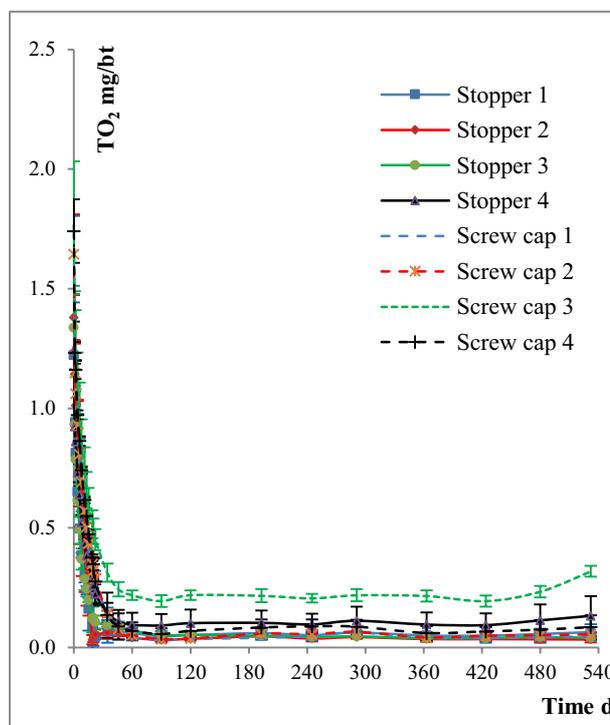


Figure 1. TO₂ = f(t) for each procedure. TO₂ mg/bt: Total oxygen in mg/bottle. Averages and standard deviations are based on 3 bottles per procedure.

due to HSO level (higher than 0.1 mg/bt at 532 d), while the DO level was equivalent to other procedures (around 0.01–0.02 mg/bt). Beyond the 90th day, we could consider that all closures reached their steady states (Fig. 1). The TO₂ stabilized at an equilibrium value which was the resultant of O₂ ingresses by the closure and O₂ consumption by the wine, for the same wine and in the same storage conditions. Thus we could link the closure OTR to the average TO₂ between 90th and 532th day (Tables 2 and 3).

The carbon dioxide concentration remained stable up to 532 days, except for the B4 procedure for which the

losses represented 18% corresponding to a loss of 60 mg/L, less than the sensory perception threshold.

3.3. Colour

The distance between two colours (ΔE_{ab}^*) was used to summarise the evolution of the colour of wine between T0 and 538 d.

$$\Delta E_{ab}^* = \sqrt{(L_0 - L_{538d})^2 + (a_0 - a_{538d})^2 + (b_0 - b_{538d})^2} \quad (1)$$

Over time, b^* increased and a^* decreased, colour tended gradually to tile colour. After 538 d, the ΔE_{ab}^* varied between 6.6 and 8.3 regardless of the procedure. C4, B1, B3 and C2 were the procedures whose colour has changed the least unlike B4 and C3. However, even if the ΔE_{ab}^* between T0 and 370 d (538 d) were at least 4.8 (6.6), the maximum interprocedural deviation was 1.7 ($\Delta E_{ab}^* = 8.3-6.6$ respectively for B4 – C4 at 538 d). The evolution of colour of wine until 538 d is mainly due to the aging of wine. By comparison, the ΔE_{ab}^* after 18 months storage is less than 1 between wines of Cabernet Sauvignon sealed by natural cork, synthetic closure and screw cap with 16 mL of headspace volume [26].

Meanwhile, the colour of red wine stored at 7 °C changed very little because $\Delta E_{ab}^* = 0.6$ and 1.1 respectively at 370 and 538 d.

And so until 18 months, the temperature impact on the colour is clearly greater than this of stopper.

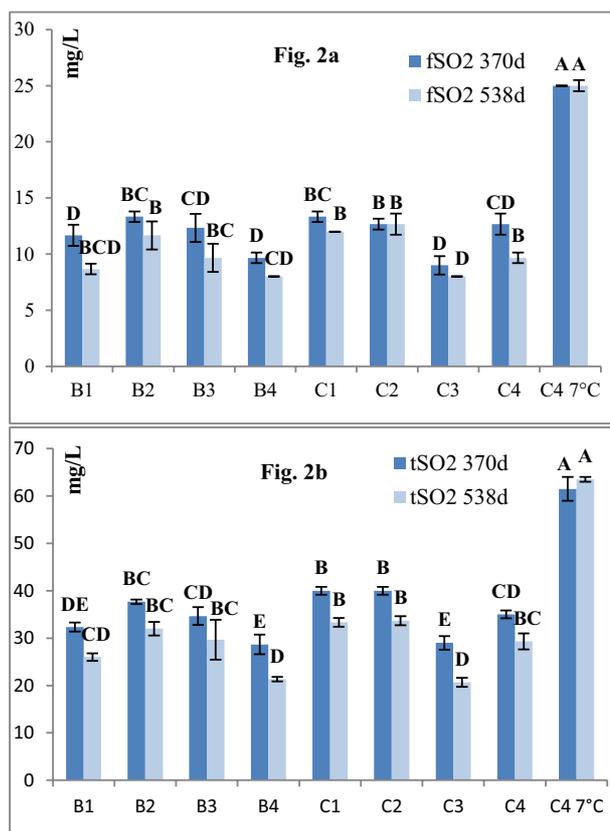
3.4. Sulfites

From 370 d, the dispersion stayed or extended between the C1, C2, B2 procedures for which the free SO₂ was at least 11 mg/L and the B4 and C3 procedures for which the free SO₂ toggled below the threshold of 10 mg/L; the B3, C4 and B1 procedures exhibiting an intermediate position (Fig. 2a). The distribution of stoppers is the same for total SO₂ (Fig. 2b).

However it should be mentioned that the impact of the stopper on the free SO₂ conservation reached a maximum of 5 mg/L between the procedures at 538 days (Fig. 2a) that is to say below the value of 7 mg/L of the expanded uncertainty for free SO₂ content of 10 to 30 mg/L [32]. Only the cold treatment had a clear effect on the preservation of free SO₂, as SO₂ consumption reactions slowed (free SO₂ at 538 d = 10/25 mg/L for C4/C4 7 °C).

3.5. Sulfite versus oxygen consumption

Oxidation of phenolic compounds leads to the production of quinones and hydrogen peroxide. SO₂ reacts with the latter, thus preventing the oxidation of ethanol according to the Fenton reaction, and reduces quinones towards their initial phenolic form. Under ideal experimental conditions, the O₂:SO₂ molar ratio of the reaction is 1:2 [33], corresponding to a maximum theoretical consumption of 4 mg of SO₂ per mg of consumed O₂. During storage of wine in bottle, a mass ratio below 4 or a molar ratio of 1 :< 2 means that a part of the oxygen which enters inside the bottle does not indirectly reacts with SO₂, but reacts with other constituents of wine [33,34]. Nucleophilic compounds come into competition with sulfites to react



Figures 2a and 2b. Free SO₂, total SO₂ at 370 and 538 days. Stoppers B1 to B4; screw caps C1 to C4. Different labels (A, B, C, D, E) indicate means that significantly differ at $p < 0.0001$. Averages and standard deviations are based on 3 bottles per procedure. fSO₂: free SO₂; tSO₂: total SO₂ in mg/L; d:day. Free and total SO₂ at T0 = $27 \pm 0/68 \pm 0$ mg/L.

with the quinones. Waterhouse et al. [35] uses the mass ratios with free SO₂ consumption (fCSO₂) and total SO₂ consumed (tCSO₂) to evidence this phenomena: more the mass ratio is less than 4 and more the oxidation of other wine constituents is important.

$$TCO_i \frac{mg}{L} = (T_{O2T0} - T_{O2i}) \frac{mg}{bt} / 0,75 + (OTR \times idays) \quad (2)$$

Free Consumed SO₂/TCO =

$$\frac{fCSO_2 \frac{mg}{L}}{TCO \frac{mg}{L}} idays = \frac{fSO_2 T0 - fSO_2 i}{TCO_i} \quad (3)$$

Total Consumed SO₂/TCO =

$$\frac{tCSO_2 \frac{mg}{L}}{TCO \frac{mg}{L}} idays = \frac{tSO_2 T0 - tSO_2 i}{TCO_i} \quad (4)$$

The ranking of stoppers in descending order of fCSO₂/TCO at 538 d (Figure 3) led to the following classification similar to that of OTR: B3 > B2 > C1, C2 > C4 > B1 ≫ B4 > C3. This classification was identical to the ratio calculated with tSO₂C at 370 and 538 days. The only difference was the slightly inverted order between C1 and C2 at 370 days for the ratio calculated with fSO₂C.

Table 4. Ranking comparison OTR vs physicochemical parameters at 538 days. Stoppers B1 to B4; screw caps C1 to C4. OTR in mg/day/closure; TO₂ at equilibrium in mg/bottle; fCSO₂ and TCO in mg/L; d: day.

Parameter	Range	OTR and impact of oxygen exposure		
		low	medium	high
OTR mg/d	0.005 to 0.067	B2, B3, C1, C2	< C4 < B1	<< B4 << C3
TO ₂ at equilibrium mg/bt	0.04 to 0.23	B2, C1 < B3, C2	< B1 < C4	<< B4 << C3
% losses free SO ₂ 538d	53 to 70	C2, C1, B2	<B3, C4 < B1	< B4, C3
ΔE* 538d	6.6 to 8.2	C4 < B1 < C2, B3	< C1, B2	< C3 < B4
fCSO ₂ /TCO 538d	3.9 to 0.5	B3, C1, C2, B2	> C4 > B1	>> B4, C3

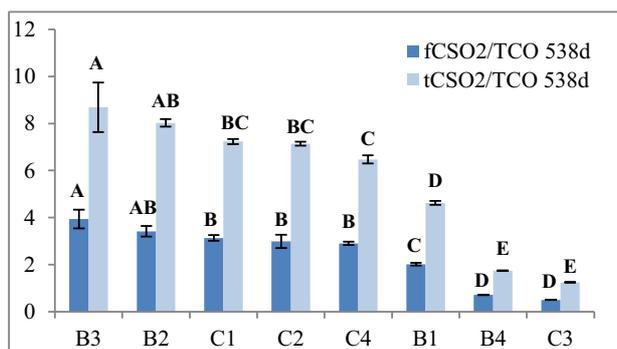


Figure 3. Ratios of free/total Consumed SO₂ / TCO at 538 days. Stoppers B1 to B4; screw caps C1 to C4. f(t)CSO₂/TCO: free (total) Consumed SO₂ / Total Consumed Oxygen. fCSO₂, tCSO₂ and TCO expressed in mg/L. Ratios ranked in descending order of fCSO₂/TCO 538 d. Different labels (A, B, C, D, E) indicate means that significantly differ at p<0.0001. Averages and standard deviations are based on 3 bottles per procedure.

Logically between these two dates, a decline was observed for all values, highlighting a more intense oxidation over time. As illustrated in Fig. 3, the ratio tCSO₂/TCO was greater than 4 for the 6 least permeable stoppers. This result has already been observed in previous studies on red wine rich in tannins after 12 and 15 months of storage evidencing oxygen-independent SO₂ consumption reactions [6,36].

TO₂ at T₀ was between 1.22 and 1.75 mg/bt according the procedures and average TO₂ of corked bottles was 1.30 mg/bt while it was 1.69 mg/bt for capped bottles (Table 3). Therefore the caps procedures started with an average handicap of 0.39 mg/bt compared to the stopper procedures linked to the bottling conditions but independent of the kind of closure. This bias arbitrarily increased the TCO of capped bottles and impacted their sulfites contents without leading the possibility to really quantify it afterwards.

In addition, the OTR of B2 included an estimated (but unmeasured) release of 1.5 mg/stopper. If we considered a release of 2 mg/stopper, mass ratios with fSO₂ and tSO₂ 538 d decreased respectively to 3.07 and 7.22, bringing B2 behind C2 and even behind C4 if we considered the handicap on TO₂ at T₀ (but excluding the unquantifiable impact on sulfites).

To the more or less accurate assessment of the release of stoppers (in this study, particularly for B2), we must add the high uncertainty on the measurement of free and total SO₂ regardless of the analytical method [32].

Finally, all these sources of variation and uncertainty influenced the ratio values and stoppers ranking.

3.6. Impact of the OTR on physicochemical parameters

TO₂ at equilibrium, fSO₂ losses and ΔEab* are physicochemical parameters which are not statistically related to the OTR, but their evolution is influenced by the diffusion of oxygen through the stopper. Table 4 collects these parameters by comparison to OTR and fCSO₂/TCO ratio at 538 d.

Table 4 showed that the impact of oxygen exposure on the red wine followed the rise of closure OTR. But more than a ranking, it rather evidenced an opposition between B2, B3, C1, C2 and B4, C3; C4 and B1 having an intermediate position.

A matter of fact, 2 mg/L of TO₂ at bottling corresponds to one year of oxygen ingresses by B3, B2, C1 or C2 closures and 5 mg/L corresponds to one year of oxygen ingresses by B1 stopper. In addition, previous studies [37,38] on wines stored at ambient temperature show that oxidised characters begin to appear around 12 mg/L (9 mg/75 cL) of oxygen ingresses in the bottle. Thus, given the concerned quantities of oxygen, it is essential to control and manage oxygen at packaging by reducing the targeted amount of total O₂ trapped in bottle (TO₂ at T₀) but also its heterogeneity (standard deviation of each lot), so that stopper fully plays its role of oxygen diffuser, especially for wines aimed to be drunk within 18 months.

3.7. Sensory analyses

For each analysis time point, jury performances were checked. The panel repeatability and consensus were good.

At 10 months, wines are significantly discriminated by the attribute *colour intensity* with pvalue <0.0001 (Fig. 4). No olfactory and taste differences were observed between the eight procedures. C2 was significantly different from all other procedures by a lighter colour. Then between the seven other wines, the C3 procedure had a darker colour than B2.

At 17 months, two olfactory descriptors (*animal*, pvalue = 0.030 and *pepper*, pvalue = 0.032) and the visual descriptor (*colour intensity*, pvalue <0.0001) allowed to discriminate wines (Fig. 4). No taste difference was observed. At olfactory level, we could observe that C2 and B2 procedures had significantly more intense animal odour than B4. The B2 procedure also had more peppery odour than B1. As at 10 months, C3 procedure had a significantly darker colour particularly than C2, B2 and C1.

The evolution of wines in 17 months differed according to the closures. The sensory evolution of wine closed by the four screw cap procedures was important, wines became less bitter and the smell of pepper decreased.

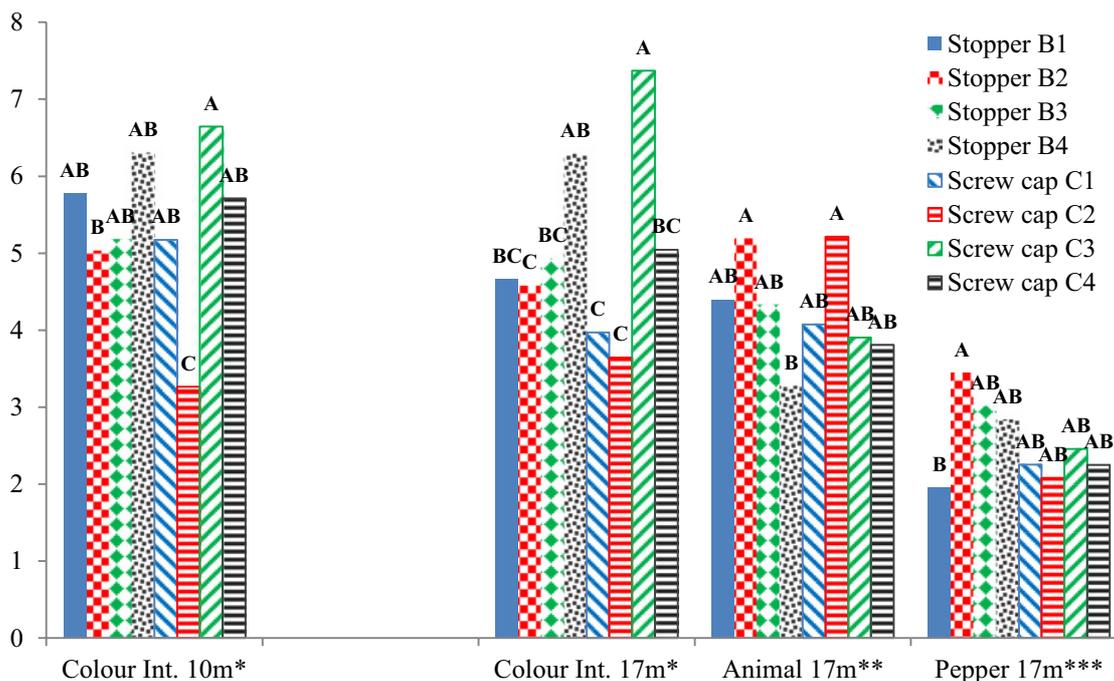


Figure 4. Means for the significant attributes at 10 and 17 months of storage. Colour Int.: Colour Intensity; m: month. Different labels (A, B, C) indicate means that significantly differ at * $p < 0.0001$; ** $p < 0.030$; *** $p < 0.032$.

However, the four stoppers procedures had different developments. For B1 and B4 procedures, animal odour decreased. On the contrary, the animal odour of B3 became more intense. About the B2 procedure, the colour of wine became lighter and the intensities of *cooked red fruit* and *pastry* decreased.

A graduation of OTR was observable on the basis of *animal* and *colour intensity*. Wines with low OTR closures had a more intense animal odour and lighter colour; those with high OTR were darker colour. This observation was made in a previous study on a Grenache wine [5] where it was shown that visual and olfactory differences were observed according to the OTR levels (max. OTR difference: 4.05 mg/year /37.5 cL of wine) but with little impact on taste attributes.

The results obtained by Kwiatkowski et al. [26] with a synthetic cork and a small headspace corroborated our conclusions: a Cabernet Sauvignon wine evolved towards significantly reduced notes, even if these descriptors were not the dominant characteristics of the wines. However, for Ugliano et al. [39], an intake of 1 mg/O₂/year was enough to modify the olfactory characteristics from reduced to fruity; but this conclusion was very dependent on the type of red wine.

4. Conclusion

As regards the physicochemical analyses until 18 months, the important points were:

- the average CO₂ losses were negligible,
- the free SO₂ was always above 10 mg/L, except for B4 and C3 closures,
- HSO of B4 and C3 closures stabilised at higher content, while there was not significant difference of DO between all the closures,

- the evolution of the colour was more and more clear with the time ($\Delta E_{ab}^* \geq 4.8$ as from 370 d), but it was more due to the aging of wine than to the impact of closure,
- the beneficial effect of storage at 7 °C for C4 screw cap was clear on SO₂ conservation and protection of the initial colour.

Finally, the ranking of closures from the overall experiment was similar to this of OTR manufacturers, namely from less to more permeable:

$$C1, C2, B2, B3 > C4 > B1 \gg B4 > C3$$

C1, C2, B2 and B3 closures were difficult to differentiate, as the oxygen ingress of the first year is around 1.8–1.9 mg for these four closures. The difficulty of finding a link between physicochemical and sensory results mostly came from the fact that between 10 and 18 months, differences of oxygen intakes were low between the stoppers (except for B4 and C3) and wines remained covered by free SO₂. But even with the most permeable closures (C3 and B4), the wines were not systematically characterized by oxidation or aging descriptors.

The study was also conducted on a Colombard and a Rosé de Provence wines under the same conditions of bottling and storage. The physicochemical analyses also highlighted the outlined OTR gradient, but it was mainly on Merlot/Tannat that sensorial analyses were affected by OTR gradient at 17–18 months (data not shown).

The authors thank the CARENE section of UNSCV for its financial support. The authors thank also for their contribution: M. Angènieux, F. Boussuge, F. Dell’Ova, E. Garcia, Y. Sire,

M. Toussaint, M. Veyret from INRA UEPR and E. Picou from INRA UMR SPO.

References

- [1] A.L. Waterhouse, V.F. Laurie, *Am. J. Enol. Vitic.* **57**, 306–313 (2006)
- [2] J.C. Danilewicz, J.T. Secombe, J. Whelan, *Am. J. Enol. Vitic.* **59**(1), 128–136 (2008)
- [3] J.C. Danilewicz, P.J. Wallbridge, *Am. J. Enol. Vitic.* **61**(1), 166–175 (2010)
- [4] P. Godden, L. Francis, J. Field, M. Gishen, A. Coulter, P. Valente, P. Høj, E. Robinson, *Aust. J. Grape Wine Res.* **7**(2), 64–105 (2001) doi: 10.1111/j.1755-0238.2001.tb00196.x
- [5] S. Caillé, A. Samson, J. Wirth, J.B. Diéval, S. Vidal, V. Cheynier, *Anal. Chim. Acta* **660**(1–2), 35–42 (2009) doi: 10.1016/j.aca.2009.11.049
- [6] M. Ugliano, J.B. Diéval, T.E. Siebert, M. Kwiatkowski, O. Aagaard, S. Vidal, E.J. Waters, *J. Agric. Food Chem.* **60**(35), 8561–8570 (2012) doi: 10.1021/jf3014348
- [7] J.C. Vidal, M. Moutounet, *OENO One* **40**(1), 35–45 (2006) doi: 10.20870/oeno-one.2006.40.1.884
- [8] V. O'Brien, C. Colby, with M. Nygaard, *Wine Industry Journal* **24**(1), 24–29 (2009)
- [9] N. Kontoudakis, P. Biosca, R. Canals, F. Fort, J.M. Canals, F. Zamora, *Aust. J. Grape Wine Res.* **14**(1), 116–122 (2008) doi: 10.1111/j.1755-0238.2008.00013.x
- [10] M. Squarzone, S. Limbo, P. Luciano, *Ind. Bevande* **XXXII**, 113–116 (2004)
- [11] A. Mas, J. Puig, N. Lladoa, F. Zamora, *J. Food Sci.* **67**(3), 1374–1378 (2002) doi: 10.1111/j.1365-2621.2002.tb10292.x
- [12] C. Puech, S. Vidal, J.F. Pegaz, C. Riou, P. Vuchot, *Rev. des Œnologues* **121**, 13–16 (2006)
- [13] G.K. Skouroumounis, M.J. Kwiatkowski, I.L. Francis, H. Oakey, D.L. Capone, B. Duncan, M.A. Sefton, E.J. Waters, *J. Aust. J. Grape Wine Res.*, **11**(2), 369–377 (2005) doi: 10.1111/j.1755-0238.2005.tb00036.x
- [14] ASTM-Standard-F1927, ASTM Internationals, West Conshohocken, PA
- [15] J. Sanchez, J.M. Aracil, *Bulletin de l'O.I.V.* **71**, 805–806 (1998)
- [16] J.B. Diéval, S. Vidal, O. Aagaard, *Packag. Technol. Sci.* **24**(7), 375–385 (2011) doi: 10.1002/pts.945
- [17] J.C. Vidal, B. Guillemat, C. Chayvialle, *Bulletin de l'OIV* **84**, 962–964 (2011)
- [18] P. Lopés, C. Saucier, C., Y. Glories, *J. Agric. Food Chem.* **53**(18), 6967–6973 (2005) doi: 10.1021/jf0404849
- [19] M. Poças, B. Ferreira, J. Pereira, T. Hogg, *Packag. Technol. Sci.* **23**, 27–33 (2009) doi: 10.1002/pts.876
- [20] P. Lopés, C. Saucier, P.L. Teissedre, Y. Glories, *J. Agric. Food Chem.* **54**(18), 6741–6746 (2006) doi: 10.1021/jf0614239
- [21] T. Karbowiak, R.D. Gougeon, J.B. Alinc, L. Brachais, F. Debeaufort, A. Voilley, D. Chassagne, *Crit. Rev. Food Sci.* **50**(1), 20–52 (2009) doi: 10.1080/10408390802248585
- [22] C. Macku, K. Reed, *Practical Winery & Vineyard Journal*, **winter 2011** (2011)
- [23] M. Brajkovich, N. Tibbits, G. Peron, C.M. Lund, S.I. Dykes, P.A. Kilmartin, L. Nicolau, *J. Agric. Food Chem.* **53**(26), 10006–10011 (2005) doi: 10.1021/jf0512813
- [24] P. Chatonnet, D. Labadie, *Rev. des Œnologues* **106**, 13–20 (2003)
- [25] P. Lopés, M.A. Silva, A. Pons, T. Tominaga, V. Lavigne, C. Saucier, P. Darrier, P., P.L. Teisseidre, D. Dubourdiou, *J. Agric. Food Chem.* **57**(21), 10261–10270 (2009) doi: 10.1021/jf9023257
- [26] M.J. Kwiatkowski, G.K. Skouroumounis, K.A. Lattey, E.J. Waters, *Aust. J. Grape Wine Res.* **13**(2), 81–94 (2007) doi: 10.1111/j.1755-0238.2007.tb00238.x
- [27] J.C. Vidal, E. Devic, F. Dell'Ova, *35th World Congress of Vine and Wine*, Izmir, 20 June (2012)
- [28] J.C. Vidal, *Rev. Fr. Œnol.* **272**, 2–5 (2015)
- [29] J.C. Vidal, M. Moutounet, *Internet journal of viticulture and enology* **4/3**, 1–15 (2011) www.infowine.com
- [30] M. Toussaint, J.C. Vidal, J.M. Salmon, *J. Agric. Food Chem.* **62**(13), 2946–2955 (2014) doi: 10.1021/jf405392u
- [31] C. Dombre, J. Wirth, M. Toussaint, C. Lixon, A. Verbaere, N. Sommerer, J.C. Boulet, S. Caillé, V. Cheynier, P. Rigou, A. Samson, J.M. Salmon, J.C. Vidal, S. Marais, Y. Gerand, P. Roux, M.H. Lemaître, A. Bobe, P. Languet, P. Chalier, *Rev. des Œnologues* **158**, 47–50 (2016)
- [32] EURL Œnologues de France, Bilan des résultats des comparaisons Interlaboratoires (CIL), janv. 2006 à déc. 2015. Union des Œnologues de France (2016)
- [33] J.C. Danilewicz, *Am. J. Enol. Vitic.* **67**(1), 13–17 (2016) doi: 10.5344/ajev.2015.15069
- [34] G. Han, M. Ugliano, B. Currie, S. Vidal, J.B. Diéval, A.L. Waterhouse, *J. Sci. Food Agric.* **95**(1), 36–43 (2014) doi: 10.1002/jsfa.6694
- [35] A.L. Waterhouse, S. Frost, M. Ugliano, A.R. Cantu, A.R., B.L. Currie, M. Anderson, A.W. Chasey, S. Vidal, J.B. Diéval, H. Heymann, *Am. J. Enol. Vitic.* **67**(3), 449–459 (2016) doi: 10.5344/ajev.2016.16006
- [36] A. Gambuti, T. Siani, L. Picariello, A. Rinaldi, M.T. Lisanti, M. Ugliano, J.B. Diéval, L. Moio, *Eur. Food Res. Technol.* (2016) doi: 10.1007/s00217-016-2780-3
- [37] J.C. Vidal, M. Toussaint, J.M. Salmon, *37th World Congress of Vine and Wine*, Mendoza, **12023** (2014)
- [38] M. Ugliano, *J. Agric. Food Chem.* **61**(26), 6125–6136 (2013) doi: 10.1021/jf400810v
- [39] M. Ugliano, J.B. Dieval, S. Begrand, S. Vidal, *Rev. des Œnologues* **156**, 45–48 (2015)