Using Biferon-B for the prevention of mastitis in cows

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Abstract. The developed method for the prevention of mastitis in lactating cows by intramuscular injection of Biferon-B at doses of 5.0–10.0–20.0 ml, provided a preventive effect in 25.0–75.0 % of animals; the best effect was achieved with Biferon-B at a dose of 5.0 ml. The use of Biferon-B in clinically healthy lactating animals was accompanied by a decrease in blood levels of stab neutrophils by 58.8–65.0 %, circulating immune complexes by 23.4–62.6 %, with a higher content of segmented neutrophils by 4.8–7.8 %, monocytes by 5.6–57.1 %, lymphocytes by 4.9–12.3 %, total immunoglobulins by 5.7–14.3 %, bactericidal and lysozyme activity of blood serum by 6.4–23.1 % and 4.3–13.7 %, respectively. The phagocytic activity of neutrophils decreased by 5.6–10.9 %. With lower indicators of the intoxication index dropped by 12.8–19.1 %, the content of average molecular peptides by 2.4–22.6 %, nitric oxide by 24.5–45.4 %, MDA by 2.8–36.7 %; catalase activity increased by 2.7–12.8 % and GPx by 10.4–29.7 %.

1 Introduction

The high incidence of mastitis in cows during all functional periods causes significant economic damage to dairy farming as a result of reduced milk productivity, milk quality, premature rejection, treatment costs, diseases of newborn calves [1, 2].

In the occurrence and spread of inflammatory diseases of the mammary gland in cows, an important role is played by factors that reduce the resistance of the mammary gland and the body as a whole [3, 4].

Modern pharmaceuticals offer a wide range of veterinary drugs and methods for treating mastitis in cows. The main active ingredients of these drugs are antibiotics. However, their ubiquitous, and often unsystematic application led to a decrease in efficiency due to the development of resistant strains of microorganisms [5, 6].

Currently, an alternative to the use of an antimicrobial drug in the treatment and prevention of mastitis is the use of immunostimulating drugs, and in particular interferons obtained using recombinant protein technologies and having a wide spectrum of action due to activation of the immune system. Interferons act against many viruses and have antibacterial properties. The bacteriostatic effect is due to a significant violation of the bioenergetic processes in microorganisms due to the depletion of tryptophan, and the mediated bactericidal effect is due to the generation of nitric oxide and reactive oxygen species in macrophages [7–9]. The protective role of interferons in the body in bacterial infections is also associated with the activation of T-lymphocytes, macrophages, and natural killers that perform a protective function [10]. Taking into account the discovered and studied properties of interferons that indicate their participation in maintaining homeostasis, drugs with antiviral and immunomodulatory effects have been developed [11, 12].

Biferon-B, a drug exhibiting antiviral and immunostimulating activity, is a mixture of bovine recombinant α- and γ-interferons with species specificity and a total antiviral activity of at least 1.0 × 10³ TCID₅₀/cm². According to its pharmacological properties, it affects the natural resistance (inducer of bactericidal and lysozyme activity) and immune status (induction of cellular and humoral immunity, endogenous cytokine system) in cattle. To date, two works are known in which data were obtained on the effect of bovine recombinant α- and γ-interferons (Biferon-B) on the nature of termination of pregnancy and the condition of cows and calves after birth [13] and the effectiveness of the combined use of bovine recombinant α- and γ-interferons and aminoseletone in subclinical mastitis in cows [14].

In this regard, the study of the influence of species-specific recombinant proteins on morphobiochemical, immunological status, microbiological and cytological indicators of the secretion of udder of cows is high on the agenda and requires detailed study.

2 Materials and methods

The studies involved 64 animals of black-and-white motley breed with milk productivity in the previous lactation of 6600–7100 kg of milk. The cows were managed tethered on a straw bedding. The groups were formed from the first day after calving. Cows were divided into four groups. On the first day after calving, the animals of the experimental group I (n = 16) were injected Biferon-B 5.0 ml intramuscularly twice with an interval of 24 hours (Scientific-Production Center

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“ProBioTech”. Republic of Belarus. The animals from experimental group II (n = 16) received 10.0 ml each; those of the experimental group III (n = 16) 20.0 ml each and in group IV (control, n = 16) the injections were not used. Biological material from animals was taken before and at the end of the experiment. Clinical observations of the animals were performed during 4 months. The state of the mammary gland was evaluated three times in a week according to the results of the study of milk with a quick mastitis test (2% solution of Masttest).

Morphological blood tests were performed on a ABX Micros 60 hematological analyzer with the definition of the leukocyte formula in accordance with the Guidelines for the Diagnosis, Treatment and Prevention of Metabolic Disorders in Productive Animals [15]. Immunological indicators, including serum bacteridal activity (SBA), and serum lysozyme activity (SLA) activity, total immunoglobulins (Ig), circulating immune complexes (CIC) in blood serum (humoral serum factors of natural non-specific resistance) and leukocyte phagocytic rate (LPR), phagocytic number (PN), phagocytic index (PI), the number of T- and B-lymphocytes (cellular immunity), were determined using standard and unified methods in accordance with the Methodological Recommendations for Assessment and Correction of the Immune Status of Animals [16]. Bacteriological studies of the secretion of the udder, the study of the cultural-morphological and biochemical properties of the isolated microorganisms was carried out by generally accepted methods according to M.A. Sidorov, D.I., Skorodumov, V.B. Fedotov [17]. Statistical processing of the obtained data was carried out using Statistica 8.0 software (StatSoft Inc., USA).

3 Results and discussion

Background morphological, biochemical, immunological blood indices, bacteriological and cytological indices of the secretion of the change of animals of all groups of indices corresponded to optimal indices. We did not reveal significant differences between the groups.

The effectiveness of the use of Biferon-B is presented in Table 1. It was found that in the group of negative control for the four-month observation period, 50.0% (8 animals) fell ill, including 25.0% of subclinical ones (4 animals) and 25.0% of clinically expressed (4 animals). When prescribing Biferon-B in 5.0 ml, one animal fell ill with subclinical and clinically expressed catarrhal mastitis, which amounted to 12.5%. The incidence of animals treated with Biferon-B at 10.0 ml was 25.0%. In this group, 2 animals fell ill with subclinical and clinically expressed catarrhal mastitis.

Among animals to which a dose of 20.0 ml Biferon-B was applied, 37.5% of cows went through mastitis, including 2 cows with subclinical mastitis and 4 cows with clinically severe catarrhal mastitis.

Consequently, the monthly treatment of cows with Biferon-B in 5.0 ml helps reduce the incidence of cows with mastitis by 37.5%. A dosage of 10.0 and 20.0 ml respectively reduces the incidence by 25.0% and 12.5%.

Thus, the optimal dose of Biferon-B with its intramuscular monthly administration twice a day with an interval of 48 hours is 5.0 ml.

The results of clinical studies were confirmed by data on the study of immunological blood counts before and after the use of interferons (Table 2).

<table>
<thead>
<tr>
<th>Indicator</th>
<th>Before experiment</th>
<th>After experiment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Erythrocytes [10^12/l]</td>
<td>5.7±0.2</td>
<td>6.4±0.3</td>
</tr>
<tr>
<td>Hemoglobin [g/l]</td>
<td>103.6±2.7</td>
<td>105.8±4.5</td>
</tr>
<tr>
<td>Leukocytes [10^9/l]</td>
<td>7.6±0.6</td>
<td>8.4±0.9</td>
</tr>
<tr>
<td>Stab neutrophils [%]</td>
<td>3.4±0.3</td>
<td>4.1±0.2</td>
</tr>
<tr>
<td>Segmented neutrophils [%]</td>
<td>42.5±2.7</td>
<td>45.4±1.4</td>
</tr>
<tr>
<td>Eosinophils [%]</td>
<td>5.0±0.4</td>
<td>4.4±0.6</td>
</tr>
<tr>
<td>Monocytes [%]</td>
<td>2.1±0.3</td>
<td>3.3±0.2</td>
</tr>
<tr>
<td>Lymphocytes [%]</td>
<td>47.0±0.6</td>
<td>49.3±0.7</td>
</tr>
<tr>
<td>Total Ig [μg/ml]</td>
<td>26.8±1.4</td>
<td>30.4±1.5*</td>
</tr>
<tr>
<td>CIC [μg/ml]</td>
<td>0.428±0.02</td>
<td>0.160±0.01***</td>
</tr>
<tr>
<td>SBA [%]</td>
<td>51.7±3.0</td>
<td>63.6±1.6*</td>
</tr>
<tr>
<td>SLA [μg/ml]</td>
<td>0.344±0.03</td>
<td>0.391±0.03*</td>
</tr>
<tr>
<td>LPR [%]</td>
<td>58.9±2.1</td>
<td>65.3±2.2</td>
</tr>
<tr>
<td>PI [cells/active phagocyte]</td>
<td>3.3±0.12</td>
<td>3.8±0.14</td>
</tr>
<tr>
<td>PN [cells/phagocyte]</td>
<td>5.6±0.39</td>
<td>6.1±0.31</td>
</tr>
<tr>
<td>MMP [a.u.]</td>
<td>1.520±0.09</td>
<td>1.176±0.08***</td>
</tr>
<tr>
<td>IEI</td>
<td>11.0±0.55</td>
<td>8.9±0.53</td>
</tr>
<tr>
<td>NO [μM/L]</td>
<td>105.9±10.0</td>
<td>75.8±4.8</td>
</tr>
<tr>
<td>MDA [μM/L]</td>
<td>4.12±0.18</td>
<td>2.61±0.12**</td>
</tr>
<tr>
<td>GPX [μM/1·min]</td>
<td>14.8±0.9</td>
<td>19.2±1.2</td>
</tr>
<tr>
<td>Catalase [μM/1·min]</td>
<td>21.9±1.3</td>
<td>27.8±1.3</td>
</tr>
</tbody>
</table>

Note: *P<0.05; **P<0.01; ***P<0.001

It was found that in the blood of cows of animals of the first group (Biferon-B, 5.0 ml each), a 58.8% decrease in the number of stab neutrophils and 62.6% in circulating immune complexes (P<0.05) was noted over the observation period, with a higher content of segmented neutrophils by 6.8%, monocytes by 57.1%, lymphocytes by 4.9%, total immunoglobulins by 14.3% (P<0.05), bactericidal and lysozyme activity of blood serum by 23.1% (P<0.05) and 13.7%, respectively, of the phagocytic activity of neutrophils by 10.9%. The intoxication index lowered by 19.1% (P<0.05), the content of average molecular peptides by 22.6% (P<0.05), nitric oxide by 45.4% (P<0.05), MDA by 36.7% (P<0.001). The activity of catalase increased by 26.9% (P<0.05), and GPX by 29.7% (P<0.05).

An increase in the number of monocytes, an increase in the absorption capacity of neutrophils testifies to the

| Table 1.  Prophylactic efficacy of Biferon-B |
|-----------------|-----------------|-----------------|-----------------|
| Indicator | I group (5.0 ml) | II group (10.0 ml) | III group (20.0 ml) | IV group (negative control) |
| Number of animals | 16 | 16 | 16 | 16 |

Diseased: subclinical mastitis [%] (animals) (%) 
- 1 (6.25) | 2 (12.5) | 2 (12.5) | 4 (25.0) 
- 1 (6.25) | 2 (12.5) | 4 (25.0) | 4 (25.0) 

Prophylactic efficacy [%] 
- 37.5 % | 25.0 % | 12.5 % |
positive effect of Biferon-B on the cells causing intensification of phagocytosis. An increase in the content of lysozyme, one of the main factors of antimicrobial protection, indicates a high proliferative activity of the granulocytes, monocytes and macrophages synthesizing it, and indicates an increase in natural nonspecific resistance. The positive effect of Biferon-B on a decrease in antigenic load under the influence of recombinant interferons is evidenced by a decrease in the number of CICs, the antigen-antibody reaction products involved in maintaining homeostasis. The normalization of the lipid peroxidation – antioxidant system is indicated by a decrease in the concentration of malondialdehyde, while increasing the activity of catalase and glutathione peroxidase.

In relation to the animals of the control group, at the end of the experiment, there was a decrease in the number of circulating immune complexes by 65.4 %, nitric oxide by 16.1 %, medium-molecular peptides by 22.9 %, intoxication index by 36.6 %, with an increase in the number of lymphocytes by 2.5 %, the content of total immunoglobulins by 23.0 %, bactericidal and lysozyme activity of blood serum by 18.4 % and 16.3 % and phagocytic activity of neutrophils by 11.9 %, respectively.

Table 3. Indicators of the immunobiochemical status of cows after administration of 10.0 ml of Biferon-B

<table>
<thead>
<tr>
<th>Indicators</th>
<th>Before experiment</th>
<th>After experiment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Erythrocytes [10^12/l]</td>
<td>5.6±0.2</td>
<td>6.4±0.3</td>
</tr>
<tr>
<td>Hemoglobin [g/d]</td>
<td>100.1±3.6</td>
<td>102.0±5.7</td>
</tr>
<tr>
<td>Leukocytes [10^9/l]</td>
<td>7.7±0.7</td>
<td>8.2±0.6</td>
</tr>
<tr>
<td>Stab neutrophils [%]</td>
<td>4.4±0.8</td>
<td>1.5±0.6</td>
</tr>
<tr>
<td>Segmented neutrophils [%]</td>
<td>40.0±2.5</td>
<td>43.1±2.1</td>
</tr>
<tr>
<td>Eosinophils [%]</td>
<td>4.9±0.3</td>
<td>3.1±0.4</td>
</tr>
<tr>
<td>Monocytes [%]</td>
<td>3.6±0.1</td>
<td>3.9±0.1</td>
</tr>
<tr>
<td>Lymphocytes [%]</td>
<td>47.1±2.7</td>
<td>49.4±2.3</td>
</tr>
<tr>
<td>Total [g/d]</td>
<td>25.6±1.3</td>
<td>28.5±0.9</td>
</tr>
<tr>
<td>CIC [g/d]</td>
<td>0.518±0.03</td>
<td>0.201±0.02**</td>
</tr>
<tr>
<td>SBA [%]</td>
<td>57.1±2.7</td>
<td>65.6±1.1*</td>
</tr>
<tr>
<td>SLA [μg/ml]</td>
<td>0.417±0.03</td>
<td>0.431±0.03</td>
</tr>
<tr>
<td>LPR [%]</td>
<td>61.2±3.18</td>
<td>64.6±3.87</td>
</tr>
<tr>
<td>PI [cells/active phagocyte]</td>
<td>3.4±0.27</td>
<td>3.7±0.14</td>
</tr>
<tr>
<td>FN [cells/phagocyte]</td>
<td>5.5±0.25</td>
<td>5.8±0.22</td>
</tr>
<tr>
<td>MMP [a.u.]</td>
<td>1.146±0.06</td>
<td>1.014±0.09</td>
</tr>
<tr>
<td>IEI</td>
<td>10.2±0.08</td>
<td>8.9±0.79</td>
</tr>
<tr>
<td>NO [μM/L]</td>
<td>73.2±4.2</td>
<td>48.2±3.7</td>
</tr>
<tr>
<td>MDA [μM/L]</td>
<td>3.84±0.12</td>
<td>3.72±0.08</td>
</tr>
<tr>
<td>GpX [μM/l-min]</td>
<td>15.1±0.8</td>
<td>17.0±1.3</td>
</tr>
<tr>
<td>Catalase [μM/mg/min]</td>
<td>22.7±1.4</td>
<td>24.9±1.4</td>
</tr>
</tbody>
</table>

Note: * P<0.05; ** P<0.01; *** P<0.001

In the blood of cows of the second group (Table 3), a decrease in the number of stab neutrophils by 65.9 % (P<0.01), the content of circulating immune complexes by 61.2 % (P<0.001), nitric oxide by 34.2 % (P<0.05) , average molecular peptides by 11.5 %, endogenous intoxication index by 12.8 % was noted, with an increase in the number of segmented neutrophils by 7.8 %, lymphocytes by 4.9 %, total immunoglobulins by 11.3 % (P<0.05), bactericidal activity of blood serum by 14.9 % (P <0.05) and SLA by 4.3 %. When using bovine recombinant α- and γ-interferons of 10.0 ml, an increase in the phagocytic activity of leukocytes by 5.6 % was established throughout the entire period of the experiment, including a phagocytic index by 8.8 % and a phagocytic number by 5.5 %, indicating the activation of cellular component of immune system.

In cows of this group, the functioning of the lipid peroxidation – antioxidant system was normalized, as evidenced by a decrease in the concentration of malondialdehyde during the experiment by 1.1 %, with a simultaneous increase in the activity of catalase and glutathione peroxidase by 9.7 and 12.6 %, respectively.

In relation to the animals of the control group, at the end of the experiment, there was a decrease in the number of circulating immune complexes by 56.4 %, nitric oxide by 30.1 %, medium-molecular peptides by 22.9 %, intoxication index by 36.6 %, with an increase in the content of total immunoglobulins by 23.0 %, bactericidal and lysozyme activity of blood serum by 18.4 % and 16.3 % and phagocytic activity of neutrophils by 11.9 %, respectively.

In cows of the third group (Table 4), there was a decrease in the number of stab neutrophils in the blood by 61.3 %, circulating immune complexes by 23.4 %, nitric oxide by 24.5 %, intoxication index by 17.0 %, with an increase in the number of segmented neutrophils by 4.8 %, lymphocytes by 12.3 %, total immunoglobulins by 5.7 %, bactericidal and lysozyme activity of blood serum by 6.4 % and 9.5 %, respectively. The indicator of the cellular component of immunity, the phagocytic activity of leukocytes, slightly decreased by 3.9 % in relation to the initial level. The phagocytic number and phagocytic index had a similar tendency.

Table 4. Indicators of the immunobiochemical status of cows after administration of 20.0 ml of Biferon-B

<table>
<thead>
<tr>
<th>Indicators</th>
<th>Before experiment</th>
<th>After experiment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Erythrocytes [10^12/l]</td>
<td>5.8±0.2</td>
<td>6.1±0.3</td>
</tr>
<tr>
<td>Hemoglobin [g/d]</td>
<td>100.6±3.2</td>
<td>103.6±9.5</td>
</tr>
<tr>
<td>Leukocytes [10^9/l]</td>
<td>8.2±1.1</td>
<td>8.4±0.6</td>
</tr>
<tr>
<td>Stab neutrophils [%]</td>
<td>3.1±0.6</td>
<td>3.5±0.0</td>
</tr>
<tr>
<td>Eosinophils [%]</td>
<td>54.0±0.3</td>
<td>50.4±0.0</td>
</tr>
<tr>
<td>Monocytes [%]</td>
<td>3.6±0.2</td>
<td>3.8±0.1</td>
</tr>
<tr>
<td>Lymphocytes [%]</td>
<td>48.6±0.9</td>
<td>49.1±3.3</td>
</tr>
<tr>
<td>Total Jg [g/d]</td>
<td>26.4±1.2</td>
<td>27.9±1.2</td>
</tr>
<tr>
<td>CIC [g/d]</td>
<td>0.410±0.03</td>
<td>0.314±0.019</td>
</tr>
<tr>
<td>SBA [%]</td>
<td>51.8±3.9</td>
<td>55.1±5.5</td>
</tr>
<tr>
<td>SLA [μg/ml]</td>
<td>0.327±0.03</td>
<td>0.358±0.019</td>
</tr>
<tr>
<td>LPR [%]</td>
<td>60.8±4.23</td>
<td>58.4±4.22</td>
</tr>
<tr>
<td>PI [cells/active phagocyte]</td>
<td>3.5±0.18</td>
<td>3.4±0.12</td>
</tr>
<tr>
<td>FN [cells/phagocyte]</td>
<td>5.7±0.33</td>
<td>5.6±0.34</td>
</tr>
<tr>
<td>MMP [a.u.]</td>
<td>1.319±0.09</td>
<td>1.287±0.06</td>
</tr>
<tr>
<td>IEI</td>
<td>13.5±0.8</td>
<td>11.2±0.7</td>
</tr>
<tr>
<td>NO [μM/L]</td>
<td>81.7±4.55</td>
<td>61.3±3.12</td>
</tr>
<tr>
<td>MDA [μM/L]</td>
<td>3.9±0.15</td>
<td>3.8±0.08</td>
</tr>
<tr>
<td>GpX [μM/l-min]</td>
<td>14.5±1.1</td>
<td>14.9±1.1</td>
</tr>
<tr>
<td>Catalase [μM/mg/min]</td>
<td>23.1±0.9</td>
<td>25.5±1.3</td>
</tr>
</tbody>
</table>

By the end of the experiment, the content of malondialdehyde decreased by 2.8 %, while the indices of the enzymatic link of antioxidant protection, on the contrary, increased. Thus, the activity of catalase and glutathione peroxidase increased by 10.4 % and 3.5 %, respectively, indicating a decrease in lipid peroxidation processes.
In relation to the animals of the control group, at the end of the experiment, there was a decrease in the content of nitric oxide by 12.4 %, medium molecular peptides by 15.7 %, intoxication index by 19.9 %, with an increase in the number of lymphocytes by 3.2 %, the content of total immunoglobulins by 11.5 %.

In the cows of the control group that were not subjected to drug treatment, the content of circulating immune complexes increased by 13.8 %, medium-molecular peptides by 10.6 %, and the number of monocytes decreased by 12.5 % (Table 5).

Table 5. Indicators of immunobiochemical status of cows in the control group

<table>
<thead>
<tr>
<th>Indicators</th>
<th>Before experiment</th>
<th>After experiment</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>Erythrocytes [10^12/l]</td>
<td>5.5±0.3</td>
<td>5.6±0.3</td>
</tr>
<tr>
<td>Hemoglobin [g/l]</td>
<td>100.4±2.1</td>
<td>104±2.3</td>
</tr>
<tr>
<td>Leukocytes [10^9/l]</td>
<td>7.7±0.4</td>
<td>7.8±0.4</td>
</tr>
<tr>
<td>Stab neutrophils [%]</td>
<td>2.8±0.2</td>
<td>3.1±0.2</td>
</tr>
<tr>
<td>Segment neutrophils [%]</td>
<td>37.4±1.8</td>
<td>35.9±2.3</td>
</tr>
<tr>
<td>Eosinophils [%]</td>
<td>6.6±0.3</td>
<td>7.7±0.5</td>
</tr>
<tr>
<td>Monocytes [%]</td>
<td>4.0±0.3</td>
<td>3.5±0.2</td>
</tr>
<tr>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>Lymphocytes [%]</td>
<td>49.2±0.9</td>
<td>49.8±2.9</td>
</tr>
<tr>
<td>Total Ig [g/l]</td>
<td>24.9±1.15</td>
<td>25.0±0.84</td>
</tr>
<tr>
<td>CIC [g/l]</td>
<td>0.406±0.04</td>
<td>0.462±0.029</td>
</tr>
<tr>
<td>SLA [μg/ml]</td>
<td>5.6±1.9</td>
<td>55.4±2.4</td>
</tr>
<tr>
<td>LPR [%]</td>
<td>0.196±0.02</td>
<td>0.252±0.037</td>
</tr>
<tr>
<td>PN [cells/ phagocyte]</td>
<td>5.6±0.21</td>
<td>5.7±0.25</td>
</tr>
<tr>
<td>MMP [a.u.]</td>
<td>1.380±0.13</td>
<td>1.526±0.06</td>
</tr>
<tr>
<td>NO [μM/l]</td>
<td>76.9±6.8</td>
<td>78.9±4.1</td>
</tr>
<tr>
<td>MDA [μM/L]</td>
<td>3.87±0.12</td>
<td>3.86±0.11</td>
</tr>
<tr>
<td>GPx [μM/min]</td>
<td>14.0±1.1</td>
<td>14.5±1.1</td>
</tr>
<tr>
<td>Catalase [μM/mg prot/min]</td>
<td>22.5±1.2</td>
<td>22.1±1.2</td>
</tr>
</tbody>
</table>

Table 6. Indicators of the secretion of mammary gland after application of Biferon-B

<table>
<thead>
<tr>
<th>Indicators</th>
<th>Before experiment</th>
<th>After experiment</th>
</tr>
</thead>
<tbody>
<tr>
<td>5.0 ml</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lysozyme [mg/ml]</td>
<td>0.323±0.020</td>
<td>0.142±0.019**</td>
</tr>
<tr>
<td>CIC [g/l]</td>
<td>0.271±0.04</td>
<td>0.132±0.008**</td>
</tr>
<tr>
<td>Total Ig [g/l]</td>
<td>2.2±0.1</td>
<td>1.7±0.1*</td>
</tr>
<tr>
<td>SC [10^9/ml]</td>
<td>73.8±13.8</td>
<td>100.6±21.6</td>
</tr>
<tr>
<td>10.0 ml</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lysozyme [mg/ml]</td>
<td>0.293±0.03</td>
<td>0.164±0.025</td>
</tr>
<tr>
<td>CIC [g/l]</td>
<td>0.205±0.04</td>
<td>0.152±0.017</td>
</tr>
<tr>
<td>Total Ig [g/l]</td>
<td>2.3±0.08</td>
<td>1.9±0.08</td>
</tr>
<tr>
<td>SC [10^9/ml]</td>
<td>93.2±20.9</td>
<td>189.8±19.4</td>
</tr>
<tr>
<td>20.0 ml</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lysozyme [mg/ml]</td>
<td>0.273±0.02</td>
<td>0.188±0.031</td>
</tr>
<tr>
<td>CIC [g/l]</td>
<td>0.258±0.04</td>
<td>0.174±0.001*</td>
</tr>
<tr>
<td>Total Ig [g/l]</td>
<td>2.3±0.1</td>
<td>2.2±0.2</td>
</tr>
<tr>
<td>SC [10^9/ml]</td>
<td>119.2±30.8</td>
<td>240.0±41.9</td>
</tr>
<tr>
<td>Negative control</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lysozyme [mg/ml]</td>
<td>0.223±0.02</td>
<td>0.345±0.019</td>
</tr>
<tr>
<td>CIC [g/l]</td>
<td>0.213±0.02</td>
<td>0.248±0.009</td>
</tr>
<tr>
<td>Total Ig [g/l]</td>
<td>1.77±0.13</td>
<td>3.42±0.34</td>
</tr>
<tr>
<td>SC [10^9/ml]</td>
<td>101.4±61.5</td>
<td>283.2±22.8*</td>
</tr>
</tbody>
</table>

The use of Biferon-B, increasing the immune status, had a positive effect on the clinical condition of lactating cows, contributing to the normalization of the immune status of the mammary gland. During the study period, in animals of the first experimental group, a decrease in the content of circulating immune complexes by 51.3 % (P<0.05), total immunoglobulins by 21.4 % (P<0.05), and lysozyme by 56.1 % (P<0.01) was observed in milk, while the content of somatic cells was not significantly changed (Table 6).

Animals of the second experimental group (10.0 ml of Biferon-B for each) showed a lower content of circulating immune complexes by 25.9 % (P<0.05), total immunoglobulins by 17.3 %, lysozyme by 44.0 %, somatic cell content increased 2.04 times (P<0.05).

In milk of cows of the third experimental group (20.0 ml Biferon-B for each), a lower content of circulating immune complexes in milk was found to be 32.6 % (P<0.05), total immunoglobulins by 4.3 %, lysozyme content by 31.1 %, however, the somatic cell content increased by 2.01 times.

Animals of the control group showed an increase in lysozyme level by 54.7 %, circulating immune complexes by 16.4 %, total immunoglobulins by 93.2 %, and somatic cells by 2.8 times.
4 Conclusion

Thus, the positive effect of the drug on the immune status of lactating cows is due to the presence of recombinant proteins in its composition, α-interferon increases the activity of natural killers, T-helpers, phagocytosis, the intensity of differentiation of B-lymphocytes, and also accelerates elimination of circulating immune complexes [17, 18].

The stimulating effect of γ-interferon is associated with the activation of macrophage phagocytic function, the production of reactive oxygen and nitrogen species, prostaglandins; in addition, it activates T-helpers and T-cytotoxic lymphocytes, stimulates the differentiation of B-cells for the production of G-immunoglobulins and the migration of lymphocytes in tissue, thereby enhancing the immune cell response [19, 20].

The studies made it possible to develop a method for the prevention of mastitis in lactating cows using Biferon-B, to show its role in optimizing the immune status of animals, reducing the number of infected animals and improving the quality of milk produced.

References

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