

Investigation of the biotechnological properties of *Zygosaccharomyces kombuchaensis* and *Gluconoacetobacter xylinus* as promising microorganisms for the production of functional drinks

Maria Babakina*, Tatyana Pershakova, and Maria Samoilenko

Krasnodar Research Institute of Agricultural Products Storage and Processing – branch of FSBSO “North-Caucasian Federal Scientific Center of Horticulture & Viniculture”, 2 st. Topolinaya alleya, Krasnodar, 350072, Russia

Abstract. The issues of biological conversion of products of processing plant raw materials remain invariably relevant. The development of processing secondary resources allows to increase the efficiency of food production, solving environmental problems and to contribute to ensuring food security. The paper presents data on the results of studies of biotechnological properties of yeast cultures *Zygosaccharomyces kombuchaensis* sp. and bacteria *Gluconoacetobacter xylinus* (Brown 1886) Yamada et al 1998, constituting a symbiotic culture called SCOBY (symbiotic culture of bacetries and yeasts), adapted and cultivated in the Krasnodar Territory with the aim of their further use for biological conversion of fruit and berry raw materials and secondary raw materials for the production of functional drinks. The optimal values for the cultivation of the studied cultures were established: pH – 6.5, temperature – for *Zygosaccharomyces kombuchaensis* sp. minimum growth temperature $t = +15-20$ °C, maximum – $t = +45-50$ °C, for *Gluconoacetobacter xylinus* (Brown 1886) Yamada et al 1998 minimum growth temperature $t = +10-25$ °C, maximum – $t = +45-50$ °C; the content of reducing substances is 10-15 %. The data obtained will be used to develop biotechnological processes for the production of functional drinks.

1 Introduction

Currently, there is a growing interest in fermented functional foods [1]. The advantage of fermentation processes is that when they are carried out, the value of unused plant waste is increased; environmental safety is ensured, since the processes are natural and do not involve the use of hazardous pollutants; efficiency of processes.

* Corresponding author: wuhdz@mail.ru

There are known researches on the technology of fermentation of sucrose, tea leaves, as well as other raw materials (cereals or plant leaves, fruit pomace, juices, coconut water) or animal products (milk, whey) [2-4].

This fermentation is carried out by a characteristic consortium of yeast and bacteria called SCOBY (symbiotic culture of bacteria and yeast) [5, 6]. The microbiological composition of a consortium can vary depending on factors such as climate, geographic location, and the environment used for the fermentation process [7, 8].

Among the acetic bacteria that are part of the consortium, *Gluconoacetobacter xylinus* (Brown 1886) Yamada et al 1998 should be distinguished, since the cellulose that makes up SCOBY is synthesized by it [9-15].

As a result of fermentation of the yeast *Zygosaccharomyces kombuchaensis* sp. a complex containing sugars is formed; polyphenols; organic food acids; cellulose fibers; amino acids; macro- and microelements such as Cu, Fe, Mn, Ni and Zn; water-soluble vitamins such as vitamin C, B vitamins; carbon dioxide; antibiotic substances and hydrolytic enzymes.

In this regard, our interest was to study the biotechnological properties of yeast cultures *Zygosaccharomyces kombuchaensis* sp. and bacteria *Gluconoacetobacter xylinus* (Brown 1886) Yamada et al 1998, constituting a symbiotic culture called SCOBY (symbiotic culture of bacteria and yeasts), adapted and cultivated in the Krasnodar Territory with the aim of their further use for biological conversion of fruit and berry raw materials and secondary raw materials for the production of functional drinks.

2 Materials and methods

As objects of research, pure cultures of yeast and bacteria that make up a symbiotic culture of bacteria and yeasts (SCOBY), adapted and cultivated in the Krasnodar Territory, were selected.

Experimental studies were carried out using a set of indicators according to standard methods used in scientific practice.

The assessment of the resistance of microorganisms to high and low pH was carried out as follows: cultures preliminarily grown to the stationary growth phase were precipitated by centrifugation (5000 rpm, 10 minutes, 25 °C), then resuspended in a liquid nutrient medium with (final cell concentration 10^8 CFU/ml), the cells were re-precipitated by centrifugation (5000 rpm, 10 minutes, 25 °C) and resuspended in a liquid nutrient medium with pH 3.5, 6.5 and 9.5. The acidity of the medium was adjusted to the required molar concentration hydrochloric acid solution (with HCl) = 0.1 mol/L and sodium hydroxide solution of molar concentration (with NaOH) = 0.1 mol/L. The analysis of the growth of the studied microorganisms was carried out at a temperature of (27 ± 1) °C. Samples were taken from the moment the culture was set on medium, then after 6, 12, 24 hours. To determine the number of cells in the medium, the method of seeding on solid nutrient media (Koch's method) was used.

To determine the temperature of optimal growth of the studied microorganisms, they were grown on Petri dishes on MPA medium for bacteria, and Sabouraud medium for yeasts. The analysis of the growth was held at temperature conditions: (10 ± 1) °C, (15 ± 1) °C, (20 ± 1) °C, (25 ± 1) °C, (30 ± 1) °C, (35 ± 1) °C and (40 ± 1) °C, (45 ± 1) °C, (50 ± 1) °C, (55 ± 1) °C. The first control of growth was taken after 2 days, the second – after the next 3-4 days.

The assessment of the resistance of microorganisms to various concentrations of reducing substances was held by centrifugation and resuspension in a liquid nutrient medium with a concentration of reducing substances (glucose and fructose in a 1:1 ratio) of 10 %, 20 %, 30 %.

3 Results and discussion

An assessment was made of the adaptive properties of the isolated and selected crops in media with different pH.

In fig. 1 shows the growth activity of the studied cultures in a medium with a pH of 3.5.

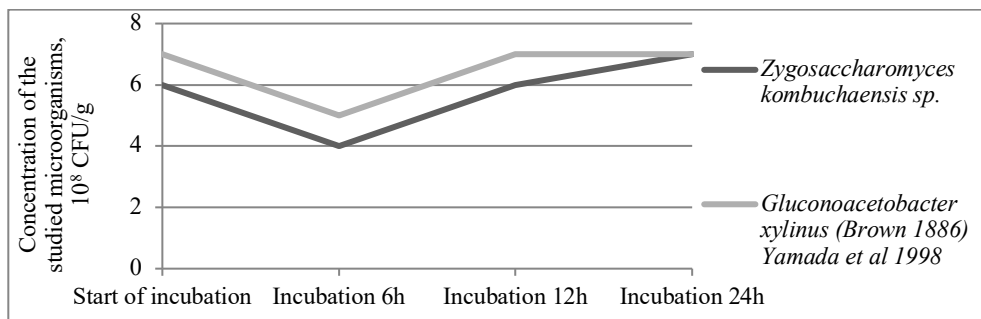


Fig.1. Growth activity of the studied microorganisms during incubation in a medium with pH 3.5

In fig. 2, you can see the growth activity of the studied cultures on a medium with a pH of 9.5.

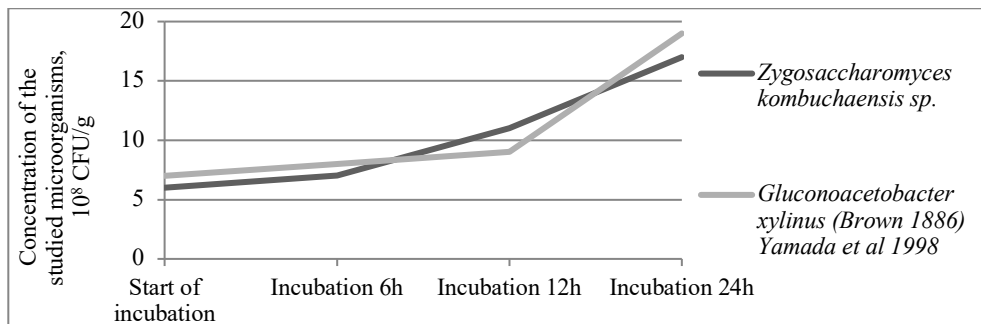


Fig. 2. Growth activity of the studied microorganisms during incubation in a medium with pH 9.5

Figure 3 shows the growth activity of the studied cultures in a medium with a pH of 6.5.

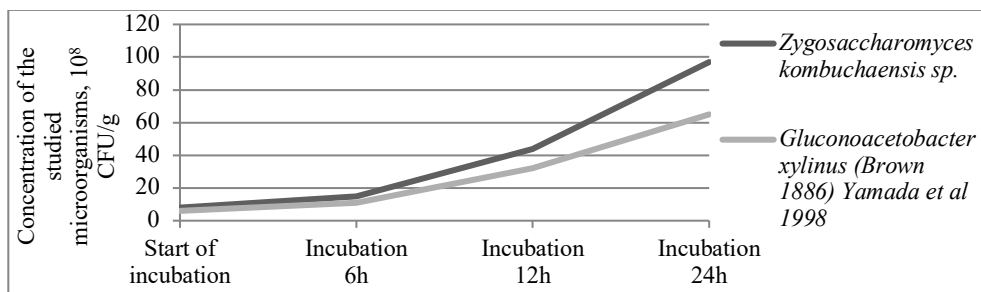


Fig. 3. Growth activity of the studied microorganisms during incubation in a medium with pH 6.5

The next step was to determine the optimal growth temperatures for the studied microorganisms. The data on the effect of temperature on the growth of the studied microorganisms are presented in Table 1.

Table 1. Influence of temperature on the growth of the studied microorganisms

Studied microorganisms	Cultivation temperature, ° C									
	10	15	20	25	30	35	40	45	50	55
<i>Zygosaccharomyces kombuchaensis</i> sp.	-	-	c	+	+	+	±	c	-	-
<i>Gluconoacetobacter xylinus</i> (Brown 1886) Yamada et al 1998	-	c	±	+	+	+	±	c	-	-

Note: "+" – active growth; "±" – less active growth; "c" – growth only after 5-6 days; "-" – no growth at all.

It was also necessary to establish the effect of the increased content of reducing substances in order to reveal the growth rate and activity of the isolated and selected microorganisms. The results are shown in Table 2.

Table 2. Survival of the studied cultures with different content of reducing substances, CFU/g

Studied microorganisms	Concentration of reducing substances, %	Incubation time, h			
		0	6	12	24
<i>Zygosaccharomyces kombuchaensis</i> sp.	10	6×10^8	17×10^8	56×10^8	130×10^8
	20	7×10^8	11×10^8	33×10^8	56×10^8
	30	5×10^8	6×10^8	10×10^8	13×10^8
<i>Gluconoacetobacter xylinus</i> (Brown 1886) Yamada et al 1998	10	5×10^8	10×10^8	38×10^8	77×10^8
	20	6×10^8	9×10^8	22×10^8	35×10^8
	30	6×10^8	5×10^8	8×10^8	11×10^8

Figure 1 shows that after 6 hours of incubation in medium with pH 3.5, the number of viable cells for each culture decreased. After 12 and 24 hours, the number of viable cells in both cultures gradually increased and returned to the initial value, 7×10^8 , but no increase in the number of cells occurred.

It can be argued that a medium with a pH of 3.5 stops and inhibits the growth of the studied microorganisms; therefore, further studies with a pH of 3.5 are inappropriate.

Based on Figure 2, it can be seen that after 24 hours of incubation, the number of *Zygosaccharomyces kombuchaensis* sp. cells increased 2.8 times from the initial one, and the number of *Gluconoacetobacter xylinus* (Brown 1886) Yamada et al 1998 cells increased 3.1 times. It can be argued that the medium with pH 9.5 does not sufficiently stimulate the growth of both studied cultures.

Based on Figure 3, it can be seen that after 24 hours of incubation, the number of cells of *Zygosaccharomyces kombuchaensis* sp. increased 12 times from the initial, *Gluconoacetobacter xylinus* (Brown 1886) Yamada et al 1998 – 9.2 times. It can be argued that the medium with pH 6.5 actively stimulates the growth of both studied cultures, therefore, it is the most optimal of all the options selected.

Based on the data in Table 1, for *Zygosaccharomyces kombuchaensis* sp. minimum growth temperature $t = +15-20$ °C, maximum – $t = +45-50$ °C. For *Gluconoacetobacter xylinus* (Brown 1886) Yamada et al 1998, the minimum growth temperature is $t = +10-25$ °C, the maximum is $t = +45-50$ °C.

Table 2 shows that the amount of yeast *Zygosaccharomyces kombuchaensis* sp. for 24 hours of cultivation in a liquid nutrient medium with reducing substances at a concentration of 10 % increased from the initial by 2066 %, in a medium with reducing substances at a concentration of 20 % – by 700 %, in a medium with reducing substances at a concentration of 30 % – by 160 %. The number of bacteria *Gluconoacetobacter xylinus* (Brown 1886) Yamada et al 1998 for 24 hours of cultivation in a liquid nutrient medium with reducing

substances at a concentration of 10 % increased from the initial by 1440 %, in a medium with reducing substances at a concentration of 20 % – by 483 %, in a medium with reducing substances in a concentration of 30 % – by 83 %.

Based on the data obtained, it can be argued that the optimal content of reducing substances in the medium for the growth of *Zygosaccharomyces kombuchaensis* sp. and *Gluconoacetobacter xylinus* (Brown 1886) Yamada et al 1998 is 10 % and should not exceed 15 %, since a further increase in the content of reducing sugars inhibits the development of both microorganisms.

4 Conclusion

To summarize, in the present study, pure cultures of *Zygosaccharomyces kombuchaensis* sp. and *Gluconoacetobacter xylinus* (Brown 1886) Yamada et al 1998, members of the symbiotic consortia, were studied under various conditions. It was found that the medium with pH 6.5 actively stimulates the growth of both studied cultures, it is the most optimal of all the options selected. The optimum concentration of reducing substances in the medium is 10-15 %. It was also revealed that both studied microorganisms actively grow at temperatures from $+(25\pm 1)^\circ\text{C}$ to $+(35\pm 1)^\circ\text{C}$.

References

1. S. Ebner, L.N. Smug, W. Kneifel, S.J. Salminen, M.E. Sanders. World J. Gastroenterol., **20(43)**, 16095-16100 (2014) <https://doi.org/10.3748/wjg.v20.i43.16095>
2. C.I. Gamboa-Gomez, L.E. Simental-Mendia, R.F. Gonzalez-Laredo, E.J. Alcantar-Orozco, V.H. Monserrat-Juarez, J.C. Ramirez-España, J.A. Gallegos-Infante, M.R. Moreno-Jiménez, N.E. Rocha-Guzmán, Food Res. Int., **102**, 690-699 (2017) <https://doi.org/10.1016/j.foodres.2017.09.040>
3. R.O. Lobo, B.K.C. Sagar, C.K. Shenoy, J. Microsop. Ultrastruct., **5**, 146-154 (2017) <https://doi.org/10.1016/j.jmau.2016.09.001>
4. A. Sknepnek, M. Pantic, D. Matijasevic, D. Miletic, S. Levic, V. Nedovic et al., Int. J. Med. Mushrooms, **20**, 243-258 (2018) <https://doi.org/10.1615/IntJMedMushrooms.2018025833>
5. S.A. Villarreal-Soto, S. Beaufort, J. Bouajila, J.P. Soucard, P. Taillandier, J. Food Sci., **83**, 580-588, (2018) <https://doi.org/10.1111/1750-3841.14068>
6. J.M. Kapp, W. Summer, Ann. Epidem., **30**, 66-70 (2018) <https://doi.org/10.1016/j.annepidem.2018.11.001>
7. S. Chakravorty, S. Bhattacharya, A. Chatzinotas, W. Chakraborty, D. Bhattacharya, R. Gachhui, Int. J. Food. Microbiol., **220**, 63-72 (2016) <https://doi.org/10.1016/j.ijfoodmicro.2015.12.015>
8. M. Coton, A. Pawtowski, B. Taminiau, G. Burgaud, F. Deniel, L. Coulloumme-Labarthe, A. Fall, G. Daube, E. Coton, FEMS Microbiol. Ecol., **93(5)**, fix048 (2017) <https://doi.org/10.1093/femsec/fix048>
9. W.N. Goh, A. Rosma, B. Kaur, A. Fazilah, A.A. Karim, R. Bhat, Int. Food. Res. J., **19**, 109-117 (2012) [http://www.ifrj.upm.edu.my/19%20\(01\)%202011/\(15\)IFRJ-2011-105%20Rajeev.pdf](http://www.ifrj.upm.edu.my/19%20(01)%202011/(15)IFRJ-2011-105%20Rajeev.pdf)
10. N. Goh, A. Rosma, B. Kaur, A. Fazilah, A.A. Karim, R. Bhat, Int. Food. Res. J., **19**, 153-158 (2012) [http://www.ifrj.upm.edu.my/19%20\(01\)%202011/\(20\)IFRJ-2011-106%20Rajeev.pdf](http://www.ifrj.upm.edu.my/19%20(01)%202011/(20)IFRJ-2011-106%20Rajeev.pdf)
11. A.-K. Waleed, K. Taous, U.-I. Mazhar, W. Fa, J. Polym. Environ., **23**, 45-53 (2015) <https://doi.org/10.1007/s10924-014-0663-x>

12. L. Wang, Z. Li, J. Hua, S. Jia, J. Zhang, H. Liu, Carbohydr. Polym., **120**, 115-119 (2015) <https://doi.org/10.1016/j.carbpol.2014.11.061>
13. M.I. Watawana, N. Jayawardena, V.Y. Waisundara, J. Food Process. Preserv., **39(6)**, 2596-2603 (2015) <https://doi.org/10.1111/jfpp.12509>
14. J.A. Viesser, G.V. de M. Pereira, D.P. Carvalho Neto, H. Rogez, A. Góes-Neto, V. Azevedo, B. Brenig, F. Aburjaile, C.R. Soccol, Int. J. Food. Microbiol., **339**, 109015 (2021) <https://doi.org/10.1016/j.ijfoodmicro.2020.109015>
15. M. Leal, V. Suárez, R. Jayabalan, H. Oros, A. Escarlante-aburto, J. Food, **16(1)**, 390-399 (2018) <https://doi.org/10.1080/19476337.2017.1410499>